



Contents lists available at ScienceDirect

Asian Pacific Journal of Tropical Biomedicine

journal homepage: www.elsevier.com/locate/apjtbReview article <https://doi.org/10.1016/j.apjtb.2017.10.016>

Plant-derived anticancer agents: A green anticancer approach

Javed Iqbal¹, Banzeer Ahsan Abbasi¹, Tariq Mahmood^{1*}, Sobia Kanwal², Barkat Ali¹, Sayed Afzal Shah¹, Ali Talha Khalil^{3,4,5}

¹Department of Plant Sciences, Quaid-i-Azam University Islamabad, 45320, Pakistan

²Department of Zoology, University of Gujrat, Sub Campus Rawalpindi, Pakistan

³Department of Eastern Medicine and Surgery, Qarshi University, Lahore, Pakistan

⁴UNESCO UNISA Africa chair in nanoscience and nanotechnology, South Africa

⁵NanoSciences African Network (NANOAFNET), South Africa

ARTICLE INFO

Article history:

Received 21 Aug 2017

Received in revised form 13 Sep 2017

Accepted 19 Oct 2017

Available online 1 Dec 2017

Keywords:

Cancer

Limitations of anticancer drugs

Phytochemicals

Analogs

ABSTRACT

Cancer is a frightful disease and represents one of the biggest health-care issues for the human race and demands a proactive strategy for cure. Plants are reservoirs for novel chemical entities and provide a promising line for research on cancer. Hitherto, being effective, chemotherapy is accompanied by certain unbearable side effects. Nevertheless, plants and plant derived products is a revolutionizing field as these are Simple, safer, eco-friendly, low-cost, fast, and less toxic as compared with conventional treatment methods. Phytochemicals are selective in their functions and acts specifically on tumor cells without affecting normal cells. Carcinogenesis is complex phenomena that involves many signaling cascades. Phytochemicals are considered suitable candidates for anticancer drug development due to their pleiotropic actions on target events with multiple manners. The research is in progress for developing potential candidates (those can block or slow down the growth of cancer cells without any side effects) from these phytochemicals. Many phytochemicals and their derived analogs have been identified as potential candidates for anticancer therapy. Effort has been made through this comprehensive review to highlight the recent developments and milestones achieved in cancer therapies using phytomolecules with their mechanism of action on nuclear and cellular factors. Furthermore, drugs for cancer treatment and their limitations have also been discussed.

1. Cancer: a global menace

Cancer is a severe metabolic syndrome and is one of the leading cause of death regardless of developments in the tools of disease diagnosis, treatment and prevention measures [1–3]. Cancer is one of the principal causes of mortality and morbidity around the globe and the number of cases are constantly increasing estimated to be 21 million by 2030 [4,5]. It is estimated that in 2017, the United States alone will have approximately 1688780 new cancer diagnoses cases and 600920 cancer deaths [6]. This uncontrolled proliferation of a normal cell which produces genetic instabilities and alterations

accumulates within cells and tissues which transforms normal cell into a malignant cell. These genetic instabilities include mutations in DNA repair genes (*p21*, *p22*, *p27*, *p51*, *p53* and tool box for DNA), tumor suppressor genes (*p53*, *NF1*, *NF2*, *RB* and biological breaks), oncogenes [*MYC*, *RAF*, *Bcl-2*, *RAS* (biological accelerators)] and genes involve in cell growth metabolism. Both external factors (radiations, smoking, tobacco, pollutants in drinking water, food, air, chemicals, certain metals and infectious agents) and internal factors (genetic mutations, body immune system and hormonal disorders) can cause cancer [7]. There are several types of cancer in human being; among these the lung cancer is reported the top listed in male followed by breast cancer in female [8,9]. Detailed information about several forms of cancer is given in Table 1. It is a major public health burden in both developing and developed countries being treated by medicinal plants as a whole or by their phytochemicals very frequently [10,11].

Previously, around 10.9 million new cancer cases, 24.6 million persons living with cancer, 6.7 million deaths reported

*Corresponding author: Tariq Mahmood, Department of Plant Sciences, Quaid-i-Azam University Islamabad, 45320, Pakistan.

Tel: +92-5190643050

E-mail: tmahmood.qau@gmail.com (T. Mahmood).

Peer review under responsibility of Hainan Medical University. The journal implements double-blind peer review practiced by specially invited international editorial board members.

around the world each year [12]. Based on World Health Organization data, above 14.1 million new cancer cases and 8.2 million deaths were mentioned globally in the year 2012 and over 70% new cancer cases has been estimated during the next twenty years [13–15]. Nearly, 80% of the world's population depend on traditional medicines and more than 60% of clinically approved anticancer drugs are derivatives of these medicinal plant [16,17]. According to literature survey, there are many anticancer drugs clinically approved and are recommended for the cancer treatment [18,19].

Among these different forms of cancer, lung cancer is reported the most in male followed by breast cancer in female. The information is gathered from the cancer stat facts (<https://seer.cancer.gov/statfacts/more.html>) and cancer statistics (2017) by Siegel.

2. Drugs for cancer treatment and their limitations

A large number of efforts have been made to minimize the harmful side effects of drugs during the process of cancer therapy like preventing the side effects on the nearby cells and tissues, increasing drug accumulation and efficacy in the lesion, developing novel drug delivery and targeting systems [20]. There are so many other methods for the treatment of cancer like they involve surgery of tumor, radiotherapy, immunotherapy, chemotherapy, cancer vaccinations, photodynamic therapy, stem cell transformation or combination thereof often accompanied by severe side effects. Such side effects include limited bioavailability, toxicity, nonspecificity, fast clearance and restriction in metastasis [21,22]. Treatment methods depend

upon the cancer type, stage and location. Chemotherapeutic agents involve cytostatic and cytotoxic drugs which have shown promising results alone or in combination with other cancer therapies. These chemotherapeutic agents involve topoisomerase inhibitors [*e.g.* irinotecan (side effects include: neutropenia, sensory neuropathy, and diarrhoea) and doxorubicin (side effects include cardiotoxicity), alkylating agents *e.g.* oxaliplatin, melphalan, carboplatin, cisplatin and cyclophosphamide (side effects include: nephrotoxicity, gastrointestinal toxicity, cardiovascular toxicity, pulmonary and hematologic toxicity), microtubules acting agent *e.g.* vincristine, vinblastine, docetaxel and paclitaxel *etc.*] [18,23]. The above mentioned drugs are highly effective against a wide range of cancers, but these drugs are also having some limitations (side effects, expensive, very complex, not eco-friendly and toxic). There are cells in our body which multiply rapidly under normal physiological conditions like hair follicle cells, bone marrow cells and digestive tract cells *etc.*, These present anticancer drugs also target these rapidly dividing normal cells which is a big challenge thus, harmful side effects arise. Due to these side effects there is decreased blood production, GIT inflammation, hair loss, immunosuppression, heart diseases and nervous disorders may arise. Another limitation is that these cancer cells resist to these drugs as they go through mutations. *e.g.*, Drug resistant genes (*ABCA4* and *ABCA12*) were over-expressed in human MCF-7 breast cancer cells respectively when docetaxel was applied. However, when phytochemical curcumin was applied in association with doce-taxel down regulation of drug resistance genes was observed [24]. Thus, treating cancer cells by employing mono-target chemical agent is not an effective method. Therefore, based on extensive research findings, phytochemicals and their derived analogues possess most promising option for the better and less toxic cancer treatment [19].

3. Current cancer therapy via phytochemicals: a novel approach

Medicinal plants serve as nature's gift to humans to help them pursue better health. Plants and their bioactive compounds are in medicinal practices since ancient times. Several medicinal plant species and their phytochemicals inhibit the progression and development of cancer [24]. It has been researched that plant kingdom comprised of approximately 250 000 plant species and only around 10% have been studied for treatment of different diseases. Phytochemicals and their derived analogues are present in different parts of the plant, *e.g.*, flower, flower stigmas, pericarp, sprouts, fruits, seeds, roots, rhizomes, stem, leaf, embryo, bark and perform several pharmacological functions. Several plant products such as alkaloids, flavonoids, lignans, saponins, terpenes, taxanes, vitamins, minerals, glycosides, gums, oils, biomolecules and other primary and secondary metabolites play significant roles in either inhibiting cancer cell activating proteins, enzymes and signaling pathways [Cdc2, CDK2 and CDK4 kinases, topoisomerase enzyme, cyclooxygenase and COX-2 (Cyclooxygenase), Bcl-2, cytokines, PI3K, Akt, MAPK/ERK, MMP, TNK, mechanistic target of rapamycin (mTOR) (detailed information in Figure 1)] or by activating DNA repair mechanism (*p21*, *p27*, *p51*, *p53* genes and their protein products), Bax, Bid, Bak proteins, stimulating the formation of protective enzymes (Caspase-3, 7, 8, 9, 10, 12), inducing antioxidant action (antioxidant enzymes

Table 1

Organ based different forms of cancers and estimated new cancer cases and deaths by 2017.

S.No.	Cancer type	Estimated new cases in 2017	Estimated deaths in 2017
1	Bladder cancer	79 030	16 870
2	Lung cancer	222 500	155 870
3	Larynx cancer	13 360	3 660
4	Non-Hodgkin lymphoma	72 240	20 140
5	Oral cavity cancer	49 670	9 700
6	Liver cancer	40 710	28 920
7	Cervical cancer	12 820	4 210
8	Kidney cancer	63 990	14 400
9	Ovary cancer	22 440	14 080
10	Endometrial cancer	61 380	19 920
11	Colon and rectum cancer	135 430	50 260
12	Anal cancer	8 200	1 100
13	Brain & nervous system cancer	23 800	16 700
14	Testis cancer	8 850	410
15	Melanoma (Skin)	87 110	9 730
16	Testis cancer	8 850	410
17	Leukemia	62 130	25 500
18	Stomach cancer	28 000	10 960
19	Prostate cancer	161 360	26 730
20	Bone and joint	3 260	1 550
21	Breast cancer	252 710	40 610
22	Oral cavity & pharynx	49 670	9 700
23	Thyroid cancer	56 870	2 010
24	Pancreas cancer	53 670	43 090
25	Small intestine	10 190	1 390
26	Hodgkin lymphoma	8 260	1 070
27	Esophagus cancer	16 940	15 690
28	Myeloma	30 280	12 590

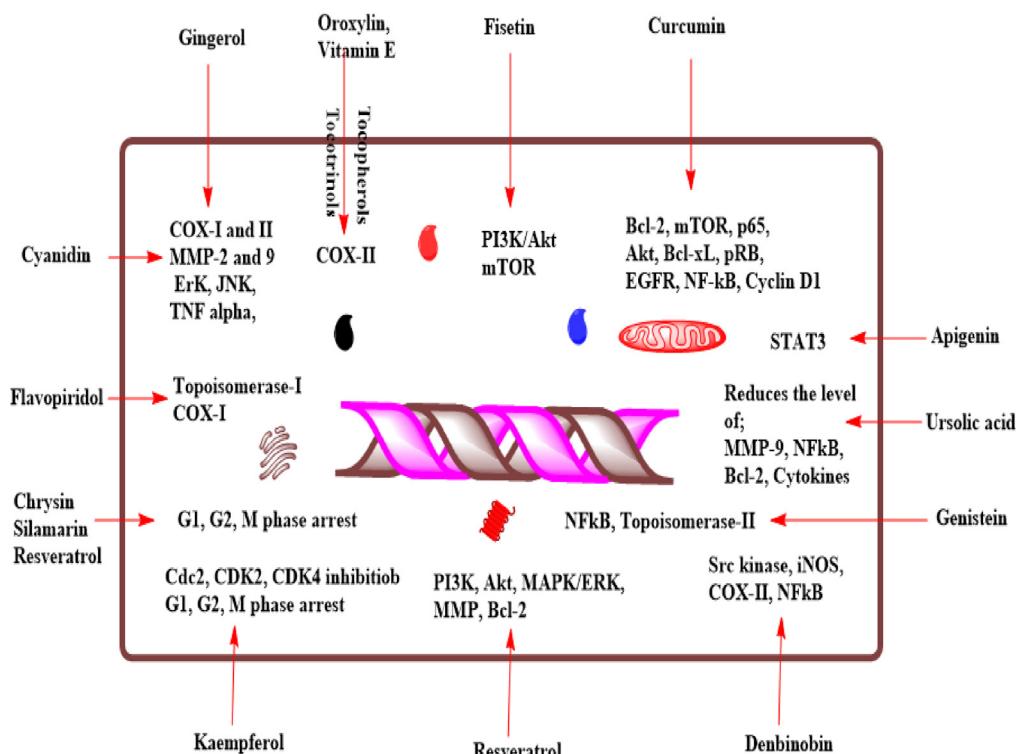


Figure 1. Impact of anticancer phytochemicals after activating expression of various genes, proteins, enzymes and signaling cascades in order to block cancer initiation and progression.

e.g. GSH, GST and GPxn), thus showing strong anticancer effects in terms of their efficacy on the above mentioned proteins, enzymes and signaling pathways (detailed information in Figure 2) [15,25]. Detailed information about these medicinal plants, family, part used and specific type of anticancer phytochemical and their mechanism of action against particular type of cancer is given in Table 2. Furthermore, the generalized model of carcinogenesis, anti-cancer mechanism of body and natural phytochemicals against cancer is discussed (Figure 3). Literature for 2010–2017 was thoroughly reviewed from ISI web of knowledge. The results are given in Figure 4.

3.1. Vinca alkaloids

Vinca alkaloids (VA) are a versatile group of phytochemicals isolated from *Catharanthus roseus* (*C. roseus*) (Apocynaceae) and are employed in the therapy of several type of cancer namely, breast, liver, leukemia, testes lung cancer. The four main VA in use are, vinorelbine, vindesine, vincristine and vinblastine [158]. The vinca alkaloids (vincristine and vinblastine) bind a specific site termed as tubulin heterodimers (vinca-binding site) disrupting the functions of microtubules or by arresting cell cycle at metaphase [159]. Currently, semi-synthetic derivatives of vinca alkaloids are vinorelbine, vindesine, vinfosiltine and vinorelbine which have been introduced in the market. These derivatives are used alone or in combination with other phytochemicals agents to fight against large number of cancers [160]. According to a scientific report, almost 64 cultivars of *C. roseus* were screened for vinca alkaloids where Cooler Rose Hot reported the highest level of serpentine alkaloids. In the near past, endophytic fungi cultured and isolated from *C. roseus* has been discovered as an alternative method for the production of the different vinca alkaloids [161].

3.2. Taxanes

Taxanes represent promising anticancer agents that act by binding to microtubules and has key role in cell division [161]. First-generation taxanes (e.g. docetaxel and paclitaxel) are strong anticancer agents in terms of their efficacy on its different molecular targets. Paclitaxel (taxol) was first extracted from the bark and leaf of *Taxus baccata* (*T. baccata*) and *T. canadensis*, *Corylus avellana* and is used to cure a wide range of cancers including ovarian, breast and lung cancer. Binding of paclitaxel with β -tubulin in the lumen of microtubules leads to decrease in microtubule dynamics and halt cell cycle at M phase while docetaxel, a semi synthetic derivative from *T. baccata* is primarily used in breast, pancreas, prostate and lung cancers therapies [161,162]. The primary mechanism of taxanes is to induce microtubule stabilization, apoptotic cell death and mitotic arrest [163]. Analogs of paclitaxel which are currently undergoing clinical trials include larotaxel, milataxel, ortataxel and tesetaxel. Larotaxel is used as alone or together with other therapies for urethral bladder, pancreatic, lung and breast cancer [164]. Furthermore, out of 2069 cancer clinical trials documented by the National Cancer Institute as of July 2004, 248 are taxane-derived drugs, containing 134 with paclitaxel, 105 with docetaxel and 10 with miscellaneous taxanes are used either alone or together with other anticancer agents [97]. Taxanes (paclitaxel, docetaxel) and its nuclear and cellular targets are given in Figure 2.

3.3. Camptothecin derivatives

Camptothecin (family of topoisomerase I poisons) is another class of plant derived clinically-active chemotherapeutic agents possesses strong anticancer potential inhibiting topoisomerase I

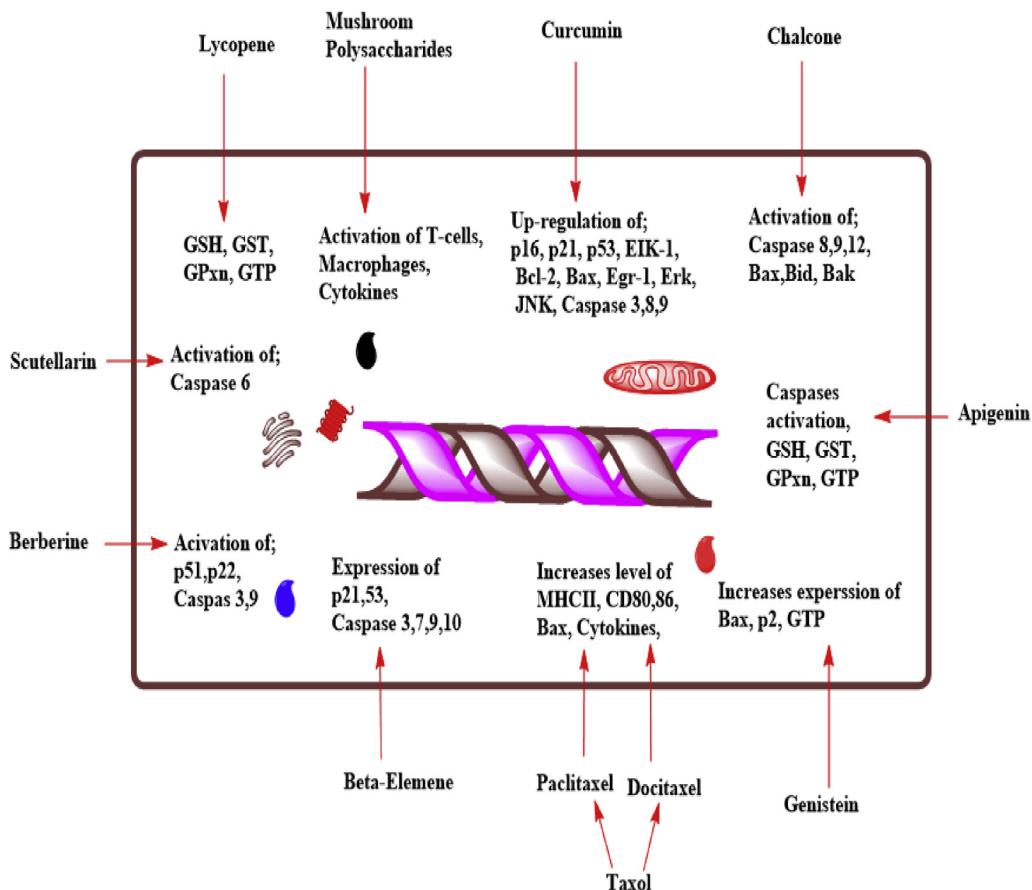


Figure 2. Impact of anticancer phytochemicals after inhibiting expression of various genes, proteins, enzymes and signaling cascades in order to block cancer initiation and progression.

in a large number of cancers [165]. It was first isolated from *Camptotheca acuminata* (Nyssaceae). The isolate of *Camptotheca acuminata* has been the only agent out of 1000 different plant extracts screened out for anticancer activity which have shown efficacy and the active constituents isolated has been identified as camptothecin. Extensive research is performed by several research organizations for effective camptothecin derivatives like topotecan (hycamtin) and irinotecan, where irinotecan is used to treat colorectal cancer while topotecan is used to treat ovarian and lung cancer [166].

3.4. *Cephalotaxus*

Cephalotaxus alkaloids are also a multipurpose group of phytochemicals that are used against wide range of cancer including A-549 lung cancer, HeLa, SGC-7901 gastric cancer cell lines. They function by inhibiting protein synthesis and targeting the molecular events in synthesis of protein such as initiation of protein synthesis, release of nascent peptide, polyribosome degradation but do not have any effect on elongation of new peptide chain [167]. Harringtonine and isoharringtonine cephalotaxus alkaloids are anticancer agents isolated from *Cephalotaxus harringtonia*. Anticancer agent like homoharringtonine has been researched to treat large number of cancers including chronic and acute myelogenous leukemia [168]. In China, harringtonine in combination with homoharringtonine are successfully used for the treatment of chronic myelogenous leukemias, acute myelogenous leukemia [16]. The homoharringtonine also been approved by FDA for the treatment of chronic myelogenous leukemia in different

countries around the world such as China, Japan, Pakistan, USA and Germany [169].

3.5. *Colchicine*

Colchicine is a natural bioactive compound isolated from *Colchicum autumnale* (Colchicaceae) and has been researched to treat several diseases like crystal arthritis, cirrhosis, gout etc. It binds permanently to tubulin, stabilizes microtubule formation, arrest cell cycle at different phases and induces apoptosis [170]. Unluckily, colchicine's action is not very specific and targets rapidly dividing normal cells and arrest their cell cycle. Therefore, semisynthetic derivatives (colchicinamide, deacetylcolchicine) of colchicine have been developed which are less toxic and are used for the treatment of variety of cancers including colorectal (HCT-116), chronic granulocytic leukemia, melanoma, central nervous system and breast cancers [50,161]. Colchicine shows toxicities and therefore is not recommended for the treatment of cancer disease. In recent years, *Gloriosa superba* in tropical regions reported to be vital source of colchicine [161].

3.6. *Ellipticine*

Ellipticine (topoisomerase II inhibitor) along with elliptine (now isoreserpiline) is also naturally occurring anticancer compound extracted from the stem, bark, leaf and root of *Bleekeria vitensis* and *Ochrosia elliptica*. These alkaloids are also found in *Aspidosperma*, *Ochrosia*, and several Apocynaceae members. The most significant DNA-damaging mechanisms of ellipticine

Table 2

List of some important medicinal plants and their phytochemicals against specific type of cancer.

Plant name	Family	Part used	Phytochemicals	Specific cancer suppressed	Literature cited
<i>Peganum harmala</i>	Zygophyllaceae	Roots	Harmine	Breast cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[26]
<i>Curcuma longa</i>	Zingiberaceae	Rhizomes	Curcumin, ascorbic acid	Leukemia, glioblastoma and colon cancer (<i>In vitro</i>)	[27]
<i>Allium wallichii</i>	Amaryllidaceae	Whole plant	Steroids, terpenoids, flavonoids, reducing sugars and glycosides	Prostate cancer, breast cancer, cervical cancer (<i>In vitro</i>)	[28]
<i>Artemisia annua</i>	Asteraceae	Whole plant	Artemisinin	Liver, breast and pancreatic cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[29]
<i>Debregeasia saeneb</i>	Urticaceae	Stem	Tannins	Internal tumors (<i>In vitro</i>)	[25]
<i>Camelia sinesis</i>	Theaceae	Leaves	Epicatechingallate, picatechin, epigallocatechin	Lung, bladder, skin, prostate and breast cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[30]
<i>Paeonia suffruticosa</i>	Paeoniaceae	Seed	Polysaccharides (HBSS, CHSS, DASS, and CASS)	Prostate, colon, human breast, and cervical cancer (<i>In vitro</i>)	[3]
<i>Ocimum sanctum</i>	Lamiaceae	Leaves	Eugenol, orientin, vicenin	Breast, liver and fibrosarcoma cancer (<i>In vitro</i>)	[31]
<i>Ginkgo biloba</i>	Ginkgoaceae	Leaves	Ginkgetin, ginkgolide A & B	Hepatocarcinoma, ovary, prostate, colon and liver cancer	[32]
<i>Camellia sinensis</i>	Theaceae	Leaves	Theabrownin	Lung cancer (<i>In vivo</i>)	[33]
<i>Ziziphus mauritiana</i>	Rhamnaceae	Leaves, bark, fruit	α -linolenic acid, Methyl stearate	Leukemia, human cervical and liver cancer (<i>In vitro</i>)	[34]
<i>Solanum nigrum</i>	Solanaceae	Leaves	Solamargine, solasonine	Breast, liver, lung and skin cancer (<i>In vitro</i>)	[35]
<i>Vigna unguiculata</i>	Fabaceae	Seeds	Black-eyed-pea trypsin/ Chymotrypsin inhibitor	Human breast cancer (<i>In vitro</i>)	[36]
<i>Ziziphus spina-christi</i>	Rhamnaceae	Flowers, leaves	Doxorubicin, spinanine-A, rutin, quercetin	Lung cancer and breast cancer (<i>In vivo</i>)	[37]
<i>Glycyrrhiza glabra</i>	Leguminosae	Roots	Licochalcone-A, licoagrochalcone	Prostate, breast, lung, stomach and kidney cancer (<i>In vivo</i>)	[38]
<i>Herba epimedii</i>	Berberidaceae	Leaves	Icarin, icaritin, icariside II	Prostate, lung, kidney and gastric cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[39]
<i>Elusine coracana</i>	Poaceae	Seeds	Ragi bifunctional inhibitor	Myeloid leukemia cell and K562 cell line (Both <i>in vitro</i> and <i>in vivo</i>)	[40]
<i>Psoralea corylifolia</i>	Leguminosae	Seeds	Psoralidin	Stomach and prostate cancer	[41]
<i>Peltophorum dubium</i>	Fabaceae	Seeds	Peltophorum dubium trypsin inhibitor	Rat lymphoma cells, human leukemia cells	[42]
<i>Vicia faba</i>	Fabaceae	Seeds	Field bean protease inhibitors	Skin cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[42]
<i>Xanthium strumarium</i>	Asteraceae	Fruit	Xanthatin	Lymphocytic leukemia and liver cancer (<i>In vitro</i>)	[43]
<i>Nigella sativa</i>	Ranunculaceae	Seeds	Thymoquinone	Colon, prostate, breast and pancreas cancer	[44]
<i>Ocimum sanctum</i>	Lamiaceae	Leaves	Eugenol, orientin, vicenin	Breast, liver and fibrosarcoma	[31]
<i>Moringa oleifera</i>	Moringaceae	Flowers, leaves	Moringa oleifera protease inhibitor (MoPI)	Abdominal cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[40]
<i>Glycine max</i>	Fabaceae	Seeds	Bowman-Birk inhibitors	Colorectal, prostate and colon cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[40]
<i>Bauhinia variegata</i>	Fabaceae	Flower	Kaempferol galactoside	Breast, lung and liver cancer (<i>In vivo</i>)	[44]
<i>Withania somnifera</i>	Solanaceae	Roots	Withaferin A, D	Breast, cervix, prostate and colon cancer (<i>In vivo</i>)	[45]
<i>Aegle marmelos</i>	Rutaceae	Bark, root	Lupeol	Lymphoma, melanoma, leukemia and breast cancer (<i>In vitro</i>)	[46]
<i>Zingiber officinale</i>	Zingiberaceae	Ginger	Gingerol	Ovary, cervix, colon, liver and urinary cancer (<i>In vitro</i> and <i>in vivo</i>)	[47]
<i>Silybum marianum</i>	Asteraceae	Flower, leaves	Silibinin	Lung, liver, skin, colon and prostate cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[48]

(continued on next page)

Table 2 (continued)

Plant name	Family	Part used	Phytochemicals	Specific cancer suppressed	Literature cited
<i>Capsicum annuum</i>	Solanaceae	Pepper	Luteolin	Colorectal cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[49]
<i>Colchicum autumnale</i>	Colchicaceae	Leaves	Colchicine	Hodgkin's lymphoma, chronic granulocytic leukemia (Both <i>in vitro</i> and <i>in vivo</i>)	[50]
<i>Aegle marmelos</i>	Rutaceae	Stem bark	Skimmianine	Liver cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[51]
<i>Boswellia serrata</i> <i>Silybum marianum</i>	Burseraceae Asteraceae	Gum Leaves, flowers	Boswellic acid Silymarin	Prostate cancer (<i>In vitro</i>) Colorectal cancer and colon cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[52] [53]
<i>Curcuma longa</i> <i>Alstonia scholaris</i>	Zingiberaceae Apocynaceae	Dried rhizome Root bark	Curcumin O-methylmacrolstonine, talcarpine, villalstonine, pleiocarpamine	Colon adenocarcinoma (<i>In vitro</i>) Lung cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[54] [55]
<i>Podophyllum peltatum</i>	Podophyllaceae	Leaves	Podophyllotoxin	Non-small cell lung carcinoma (Both <i>in vitro</i> and <i>in vivo</i>)	[56]
<i>Andrographis paniculata</i>	Acanthaceae	Whole plant	Andrographolide	Colon cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[49]
<i>Ziziphus jujuba</i>	Rhamnaceae	Fruits, seeds, leaves	Linoleic acids, triterpenoids	Breast cancer, human Jurkat leukemia T cells (Both <i>in vitro</i> and <i>in vivo</i>)	[57]
<i>Podophyllum hexandrum</i>	Berberidaceae	Leaves	Podophyllotoxin	Breast, ovary, lung, liver, bladder and testis cancer (<i>In vitro</i>)	[58]
<i>Betula utilis</i> <i>Panax ginseng</i>	Betulaceae Araliaceae	Bark Roots	Betulinic acid Panaxadiol	Melanomas (<i>In vitro</i>) Human colon cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[59] [60]
<i>Panax pseudoginseng</i> <i>Gossypium hirsutum</i>	Araliaceae Malvaceae	Roots Cotton	Panaxadiol Gossypol	Human colon cancer (Both <i>in vitro</i> and <i>in vivo</i>) Mice xenograft (HT-29) and colorectal cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[60] [61]
<i>Passiflora caerulea</i> <i>Plumbago zeylanica</i>	Passifloraceae Plumbaginaceae	Flower Leaves	Chrysin Plumbagin	Colorectal cancer (<i>in vitro</i>) Liver, fibrosarcoma, leukemia and breast cancer (<i>In vitro</i>)	[62] [63]
<i>Capsicum annuum</i>	Solanaceae	Pepper	Luteolin	Colorectal cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[49]
<i>Zingiber officinale</i> <i>Curcuma longa</i>	Zingiberaceae Zingiberaceae	Rhizomes Root, rhizome	6-Shogaol Curcumin	Ovary cancer (<i>In vitro</i>) Breast, lung, colon, prostate esophagus, liver and skin cancer (<i>In vitro</i>)	[64] [65]
<i>Oldenlandia diffusa</i>	Rubiaceae	Stem bark, leaves, fruit peel	Ursolic acid	Lungs, ovary, uterus, stomach, liver, colon, rectum and brain cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[66]
<i>Zingiber officinale</i> <i>Zingiber officinale</i>	Zingiberaceae Zingiberaceae	Ginger Root	6-Shogaol Gingerol	Ovary cancer (<i>in vitro</i>) Colon, breast and ovarian cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[64] [47]
<i>Broussonetia papyrifera</i>	Moraceae	Fruits, leaf, bark	2S-abyssinone Ilverubulin	Glioblastoma and brain cancer (<i>In vitro</i>)	[67]
<i>Glycyrrhiza uralensis</i> <i>Boerrhavia diffusa</i>	Fabaceae Nyctaginaceae	Roots Roots	Isoliquiritigenin Punarnavine	Human lung cancer (<i>In vitro</i>) Malignant melanoma cancer (<i>In vitro</i>)	[68] [69]
<i>Vitis vinifera</i> <i>Polygonum cuspidatum</i>	Vitaceae Polygonaceae	Seeds extract Whole plant	Procyanidins Resveratrol	Human colon cancer (<i>In vitro</i>) Colorectal, skin and liver cancer (<i>In vitro</i>)	[70] [71]
<i>Morinda citrifolia</i> <i>Biophytum sensitivum</i>	Rubiaceae Oxalidaceae	Roots Fruits and berries	Dammcanthal Alcoholic extract	Lung cancer, sarcomas (<i>In vitro</i>) Dalton's lymphoma ascites, Ehrlich ascites carcinoma (<i>In vitro</i>)	[72] [73]
<i>Gossypium hirsutum</i>	Malvaceae	Whole plant	Gossypol	Breast, stomach, liver, prostate and bladder cancer (<i>In vitro</i>)	[74]
<i>Aloe vera</i>	Asphodelaceae	Leaves	Alexin B, emodin	Leukemia, stomach cancer (<i>In vivo</i>)	[75]
<i>Vaccinium macrocarpon</i> <i>Annona crassiflora</i>	Ericaceae Annonaceae	Fruit Leaves	Hydroxycinnamoyl ursolic acid Caffeic acid, sinapic acid, rutin	Cervical, prostate cancer (<i>In vitro</i>) Glioma, renal, ovary cancer (<i>In vitro</i>)	[76] [77]

Table 2 (continued)

Plant name	Family	Part used	Phytochemicals	Specific cancer suppressed	Literature cited
<i>Annona coriacea</i>	Annonaceae	Seeds	Ferulic and sinapic acid	Glioma, lymphoid melanoma, lung, renal and ovary cancer	[77]
<i>Argemone gracilenta</i>	Papaveraceae	Whole plant	Argemoneine and berberine	B-cell lymphoma, leukemia (<i>In vitro</i>)	[78]
<i>Psoralea corylifolia</i>	Leguminosae	Seeds	Bavachanin, coryfolinin, psoralen	Lung, osteosarcoma, fibrosarcoma and liver cancer (<i>In vitro</i>)	[79]
<i>Moringa oleifera</i>	Moringaceae	Leaves	Niazinine A	Blood cancer (<i>In vitro</i>)	[80]
<i>Amoora rohituka</i>	Meliaceae	Stem bark	Amooranin	Lymphocytic leukemia (<i>In vitro</i>)	[81]
<i>Conyzza Canadensis</i>	Asteraceae	Roots	Conyzapyranone A and B	Epidermoid carcinoma (<i>In vitro</i>)	[82]
<i>Ziziphus rugosa</i>	Rhamnaceae	Pericarp and seed	Betulinic acid	Cytotoxicity against human melanoma cells (<i>In vivo</i>)	[83]
<i>Panax ginseng</i>	Araliaceae	Leaves	Panaxadiol, panaxatriol	Breast, ovary, lung, prostate and colon cancer (<i>In vitro</i>)	[84]
<i>C. roseus</i>	Apocynaceae	Leaves	Vinblastine, Vincristine	Breast, ovary, cervix, lung, rectum and testis cancer (<i>In vitro</i>)	[85]
<i>Centella asiatica</i>	Apiaceae	Leaves	Asiatic acid	Melanoma, glioblastoma, breast (<i>In vivo</i>)	[86]
<i>Viscum album</i>	Santalaceae	Sprouts	Viscumin, digallic acid	Breast, cervix, ovary, stomach, colon, kidney, lung cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[87]
<i>Leea indica</i>	Vitaceae	Leaves	Gallic acid	Ehrlich ascites carcinoma (<i>In vitro</i> and <i>in vivo</i>)	[88]
<i>Liriodendron tulipifera</i>	Magnoliaceae	Stem	Costunolide, tulipinolide, liriodenine, germacranoide	KB (Oral cancer), HT29 cell line (Both <i>in vitro</i> and <i>in vivo</i>)	[89]
<i>Viscum album</i>	Santalaceae	Fruits	Viscumin, digallic acid	Breast, ovary, lung, kidney, bladder and testis cancer (<i>In vitro</i>)	[87]
<i>Cicer arietinum</i>	Fabaceae	Seeds	Bowman-Birk-type protease	Breast and prostate cancer (<i>In vitro</i>)	[90]
<i>Crocus sativus</i>	Liliaceae	Dry stigmas	Crocin	Hippocampal cell death and lung cancer (<i>In vivo</i>)	[91]
<i>Centella asiatica</i>	Apiaceae	Leaves	Asiatic acid	Melanoma, glioblastoma and breast cancer (<i>In vivo</i>)	[92]
<i>Tylophora indica</i>	Asclepiadaceae	Leaves	Tylophorine	Breast cancer (<i>In vivo</i>)	[93]
<i>Dioscorea colletti</i>	Dioscoreales	Rhizomes	Dioscin	Liver and human gastric cancer (<i>In vitro</i>)	[94]
<i>Croton macrobotrys</i>	Euphorbiaceae	Leaves	Corydine, salutaridine	Leukemia and lung cancer (<i>In vitro</i>)	[95]
<i>Clausena lansium</i>	Rutaceae	Seeds	Clausenalansamid A and B	Gastric, liver cancer (<i>In vitro</i>)	[96]
<i>Bleekeria vitensis</i>	Apocynaceae	Leaf	Elliptinium	Myelogenous leukemia and breast cancer (<i>In vivo</i>)	[97]
<i>Combretum caffrum</i>	Combretaceae	Bark, kernel and fruit	Combretastatins	Colon, and leukemia and lung cancer (<i>In vivo</i>)	[97]
<i>Solanum lycopersicum</i>	Solanaceae	Fruit	Lycopene	Prostate and colon cancer (<i>In vivo</i>)	[98]
<i>Plumbago zeylanica</i>	Plumbaginacea	Roots	Plumbagin	Blood and skin cancer (<i>In vitro</i>)	[99]
<i>Crocus sativus</i>	Iridaceae	Flower stigmas	Crocin, picrocrocin, crocetin, and safranal	Sarcoma and oral cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[100]
<i>Actaea racemosa</i>	Ranunculaceae	Rhizomes, roots	Actein	Liver and breast cancer (<i>In vivo</i>)	[101]
<i>Peristrophe bicalyculata</i>	Acanthaceae	Aerial parts	β-Caryophyllene, α-zingiberene	Breast cancer (<i>In vitro</i>)	[102]
<i>Cannabis sativa</i>	Cannabinaceae	Leaf	Cannabinoid	Lung, pancreas, breast, prostate and colorectal cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[103]
<i>Silybum Marianum</i>	Asteraceae	Flower, leaves	Silymarin	Colorectal cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[104]
<i>Enterolobium contortisiliquum</i>	Fabaceae	Seeds	Enterolobium contortisiliquum trypsin inhibitor	Gastric and breast cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[105]
<i>Linum usitatissimum</i>	Linaceae	Leaves, flowers	Cynogenetic glycosides	Breast cancer (<i>In vitro</i>)	[106]
<i>Calvatia caelata</i>	Agaricaceae	Fruiting bodies	Laccases (Enzymes)	Liver, breast cancer (<i>In vitro</i>)	[107]
<i>Tylophora indica</i>	Combretaceae	Bark, Kernel fruit	Tylophorine	Breast cancer (<i>In vivo</i>)	[97]
<i>Allium sativum</i>	Amaryllidaceae	Buds, leaves	Allylmercaptocysteine, allicin	Lymphoma, cervix cancer (<i>In vivo</i>)	[108]

(continued on next page)

Table 2 (continued)

Plant name	Family	Part used	Phytochemicals	Specific cancer suppressed	Literature cited
<i>Hibiscus mutabilis</i>	Malvaceae	Pepper	Lectin	Liver, breast cancer (<i>In vitro</i>)	[109]
<i>Plumbago zeylanica</i>	Plumbaginaceae	Roots	Plumbagin	Blood cancer, skin cancers (<i>In vitro</i>)	[99]
<i>Saffron crocus</i>	Iridaceae	Dry stigmas	Saffron	Liver, lung cancer and pancreatic cancer (<i>In vitro</i>)	[110]
<i>Taxus brevifolia</i>	Taxaceae	Bark	nab-Paclitaxel	Ovarian and breast cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[23]
<i>Vitis vinifera</i>	Vitaceae	Fruit	Cyanidin	Colon cancer (<i>In vivo</i>)	[111]
<i>Actaea racemosa</i>	Ranunculaceae	Rhizomes and roots	Actein	Liver and breast cancer (<i>In vivo</i>)	[101]
<i>Pyrus malus</i>	Rosaceae	Bark, fruit	Quercetin, procyanidin	Colon cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[112]
<i>Betula Sp.</i>	Betulaceae	Leaves	Betulinic acid	Human melanoma xenografts and leukemia (<i>In vitro</i>)	[113]
<i>Tabernaemontana divaricata</i>	Apocynaceae	Leaves	Cononitarine B, Conophylline	Liver, lung, breast and colon cancer (<i>In vitro</i>)	[114]
<i>Smilax chinensis</i>	Liliaceae	Rhizomes	Tannin, saponins and flavonoid	Sarcoma-180 and ascites sarcoma (Both <i>in vitro</i> and <i>in vivo</i>)	[112]
<i>Allium sativum</i>	Liliaceae	Whole plant	Allin	Carcinoma of human (mammary) gland (Both <i>in vitro</i> and <i>in vivo</i>)	[115]
<i>Aloe vera</i>	Liliaceae	Whole plant	Aloesin, emodin	Anti-angiogenic activity (<i>In vitro</i>)	[116]
<i>Curcuma longa</i>	Zinziberaceae	Roots	Curcumin	Stomach cancer (<i>In vitro</i>)	[117]
<i>Emblica officinalis</i>	Euphorbiaceae		Polyphenols, tannins	Lymphoma and melanoma (<i>In vitro</i>)	[118]
<i>Momordica charantia</i>	Cucurbitaceae	Leaves, Roots	Charantin, cucurbitane-type triterpene	Colon cancer and breast cancer (<i>In vitro</i>)	[119]
<i>Stevia rebaudiana</i>	Asteraceae	Leaves	Labdane sclareol properties	Anti-tumorous and cytotoxic (<i>In vitro</i>)	[120]
<i>Camellia sinensis</i>	Theaceae	Leaves	Epigallocatechin gallate	Brain, prostate, cervical and bladder cancer (<i>In vivo</i>)	[121]
<i>Nelumbo nucifera</i>	Nelumbonaceae	Embryos	Neferine	Liver cancer (<i>In vitro</i>)	[122]
<i>Ocimum sanctum</i>	Lamiaceae	Leaves	Caryophyllene, camphor	Sarcoma-180 solid tumor (<i>In vitro</i>)	[11]
<i>Calvatia caelata</i>	Agaricaceae	Fruiting bodies	Calcaelin	Breast and spleen cancer cells (<i>In vivo</i>)	[123]
<i>Pleurotus sajor-caju</i>	Agaricaceae	Fruiting bodies	Ribonucleases	Leukemia and liver cancer (<i>in vivo</i>)	[124]
<i>Lentinus edodes</i>	Marasmiaceae	Fruiting bodies	Lentinan	Sarcoma-180 in mice (<i>In vivo</i>)	[124]
<i>Schizophyllum commune</i>	Schizophyllaceae	Fruiting bodies	Schizophylan	Head and neck cancer (<i>In vivo</i>)	[125]
<i>Matricaria chamomilla</i>	Asteraceae	Whole plant	Apigenin	Colorectal cancer (<i>in vivo</i>)	[126]
<i>Fagopyrum sculentum</i>	Polygonaceae	Seeds	Buckwheat inhibitor-1 protein	T-acute lymphoblastic leukemia (T-ALL) cells (<i>in vitro</i>)	[127]
<i>Glycine max</i>	Fabaceae	Seeds	Soybean trypsin inhibitor	Human ovarian cancer (<i>in vivo</i>)	[128]
<i>Ipomoea batata</i>	Convolvulaceae	Roots	Trypsin inhibitor protein	Promyelocytic leukemia cells (<i>In vitro</i> and <i>in vivo</i>)	[129]
<i>Lavatera cashmeriana</i>	Malvaceae	Seeds	Lavatera cashmeriana protease inhibitors (LC-pi I, II, III)	Leukemia, lung, colon cancer (<i>In vitro</i>)	[130]
<i>Lens culinaris</i>	Fabaceae	Seeds	Lentil (Lens culinaris trypsin inhibitor)	Human colon cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[131]
<i>Medicago scutellata</i>	Fabaceae	Seeds	Medicago scutellata trypsin inhibitor	Human breast and cervical cancer (<i>In vitro</i>)	[132]
<i>Phaseolus acutifolius</i>	Fabaceae	Seeds	Tepary bean protease inhibitor	Leukemia L1210 and lymphoma MBL2 (<i>In vitro</i>)	[133]
<i>Pisum sativum</i>	Fabaceae	Pea	Protease inhibitors, rTI1B, rTI2B	Human colorectal and colon cancer (<i>In vitro</i>)	[134]
<i>Phaseolus vulgaris</i>	Fabaceae	Seeds	Tepary bean protease inhibitor	Leukemia L1210 and lymphoma MBL2 (<i>In vitro</i>)	[133]
<i>Coccinia grandis</i>	Cucurbitaceae	Leaves	(CG) protease inhibitors	Colon cancer (<i>In vitro</i>)	[135]
<i>Ginkgo biloba</i>	Ginkgoaceae	Leaves	EGb and bilobalide	Colon cancer (<i>In vivo</i>)	[136]
<i>Curcuma zedoaria</i>	Zingiberaceae	Whole plant	Curcumin	Colorectal cancer and B-16 melanoma cells (<i>In vitro</i>)	[137]
<i>Clematis manshrica</i>	Ranunculaceae	Flower, Leaves	1,4-benzoquinone, 5-o-ethyl-embelin, 15-carbon isoprenoid	Liver cancer and blood cancer (<i>In vivo</i>)	[138]

Table 2 (continued)

Plant name	Family	Part used	Phytochemicals	Specific cancer suppressed	Literature cited
<i>Vitex agnus- castu</i>	Verbenaceae	Fruit	Vitex or luteolin	Human uterine, ovarian, cervical and breast cancer (<i>In vitro</i>)	[139]
<i>Withania somnifera</i>	Solanaceae	Root stem and leaves	Adriamycin and 5-fluorouracil	Human cervical cancer cell (<i>In vitro</i>)	[140]
<i>Aristolochia fontanesii</i>	Aristolochiaceae	Roots	Aqueous extract	Breast cancer (<i>In vitro</i>)	[141]
<i>Centella asiatica</i>	Apiaceae	Whole plant	Asiatic acid, Tamoxifen	Breast cancer (<i>In vitro</i>)	[142]
<i>Carissa spinarum</i>	Apocynaceae	Fruit	Alkaloids, saponins, tannins, flavonoids	Nasopharyngeal carcinoma (<i>In vitro</i>)	[143]
<i>Asclepias curassavica</i>	Asclepiadaceae	Aerial pats	Asclepin, cardenolides	Liver cancer (<i>In vitro</i>)	[144]
<i>Annona squamosa</i>	Annonaceae	Seed	Bullatacin	Liver cancer (<i>In vitro</i>)	[145]
<i>Bryophyllum pinnatum</i>	Crassulaceae	Leaves	Bryophyllin A	Cervical cancer (<i>In vitro</i>)	[146]
<i>Butea monosperma</i>	Fabaceae	Flower	Butrin, (7,3',4'-trihydroxyflavanone-7,3'-diglucoside)	Liver cancer (<i>In vitro</i> and <i>in vivo</i>)	[147]
<i>Vitex negundo</i>	Lamiaceae	Fruit	Chrysoplenitin	Human pancreatic cancer (<i>In vitro</i>)	[148]
<i>Moringa oleifera</i>	Moringaceae	Seed	Pterygospermin 4-(4'-O-acetyl- α -L-rhamnopyranosyloxy), benzylisothiocyanate,4-benzylisothiocyanate	Lung, neuroblastoma and colon cancer (<i>In vitro</i>)	[149]
<i>Syzygium cumini</i>	Myrtaceae	Fruit	Kaempferol-7-O-methylether, γ -sitosterol	Leukemia (<i>In vitro</i>)	[150]
<i>Argemone mexicana</i>	Papaveraceae	Leaves	Pancorine, (p)-argenaxine, (p)-higenamine, angoline	Gall bladder and breast cancer (<i>In vivo</i>)	[151]
<i>Citrus limon</i>	Rutaceae	Fruits	5-hydroxy-6,7,8,3',4'-pentamethoxyflavone	Human colon cancer (<i>In vitro</i>)	[152]
<i>Taxus wallichiana</i>	Taxaceae	Stem bark	Diterpenoid 2-deacetoxytaxinine	Breast and kidney cancer (Both <i>in vitro</i> and <i>in vivo</i>)	[153]
<i>Berberis vulgaris</i>	Berberidaceae	Roots stem and bark	Berberine	Breast, liver, colon cancers (<i>In vitro</i>)	[154]
<i>C. roseus</i>	Apocynaceae	Bark, leaves	Vindesine	Leukemias, testicular, breast and lung cancer (<i>In vitro</i>)	[113]
<i>C. roseus</i>	Apocynaceae	Bark, leaves	Vincristine	Lymphocytic leukemia (<i>In vivo</i>)	[113]
<i>C. roseus</i>	Apocynaceae	Bark, leaves	Vinblastine	Lymphocytic leukemia (<i>In vivo</i>)	[113]
<i>T. baccata</i>	Taxaceae	Bark, leaves	Cabazitaxel	Prostate cancer (<i>In vivo</i>)	[154]
<i>Colchicum autumnale</i>	Colchicaceae	Leaves	Colchicine	Multiple solid tumors (<i>In vitro</i> and <i>in vivo</i>)	[155]
<i>T. baccata</i>	Taxaceae	Bark, leaves	Larotaxel	Breast, bladder and pancreatic cancer (<i>In vivo</i>)	[156]
<i>Taxus brevifolia</i>	Taxaceae	Bark	Paclitaxel	Breast and ovarian cancer (<i>In vivo</i>)	[113]
<i>Berberis vulgaris</i>	Berberidaceae	Root, stem bark	Berberine, cannabisin	Breast, prostate and liver cancer (<i>In vivo</i>)	[157]

were considered to be topoisomerase II inhibition and intercalation with DNA in order to avoid proliferation [171]. Ellipticine and their derivatives e.g. N-2-(diethylaminoethyl)-9-hydroxyellipticinium chloride, 2-N-methyl 9-hydroxyellipticine are efficient anticancer compounds and are used to cure ependymoblastoma, leukemia, myeloma, melanoma, breast and colon cancer [172,173]. Ellipticine also perform their functions by inhibiting p53 protein phosphorylation and inhibit CDK2 kinase in human lung and colon cancer. A derivative of ellipticine (elliptinium) is also in clinical trials in France to check out its anticancer potential against breast cancer [174].

3.7. Berberine

Berberine is a strong anticancer compound in terms of its efficacy and clinical trials isolated from the root and rhizome of *Tinospora cordifolia*, *Berberis vulgaris*, *Berberis aquifolium*

and *Rhizoma coptidis* [175]. Berberine has been used for the treatment of variety of cancers namely; breast, prostate and colorectal cancer [176]. Berberine induces apoptosis and cell cycle arrest at G2/M phase in breast, colorectal and liver cancer, inhibit anti-apoptotic proteins c-IAP1 and Bcl-2, activate pro-apoptotic proteins (p21, p53, caspase-3 and caspase-9) [177]. The molecular targets of berberine are illustrated in Figure 2.

3.8. Combretastatins

The combretastatins is a class of anti-angiogenic agents isolated from *Combretum caffrum* (Combretaceae) and specifically suppresses tumor angiogenesis. For this purpose, National Cancer Institute has collaborated with botanical Research Institute of South Africa and developed novel anticancer agent combretastatins and many derivatives. Combretastatin of family Stilbenes act against tumor and causes tumor necrosis due to

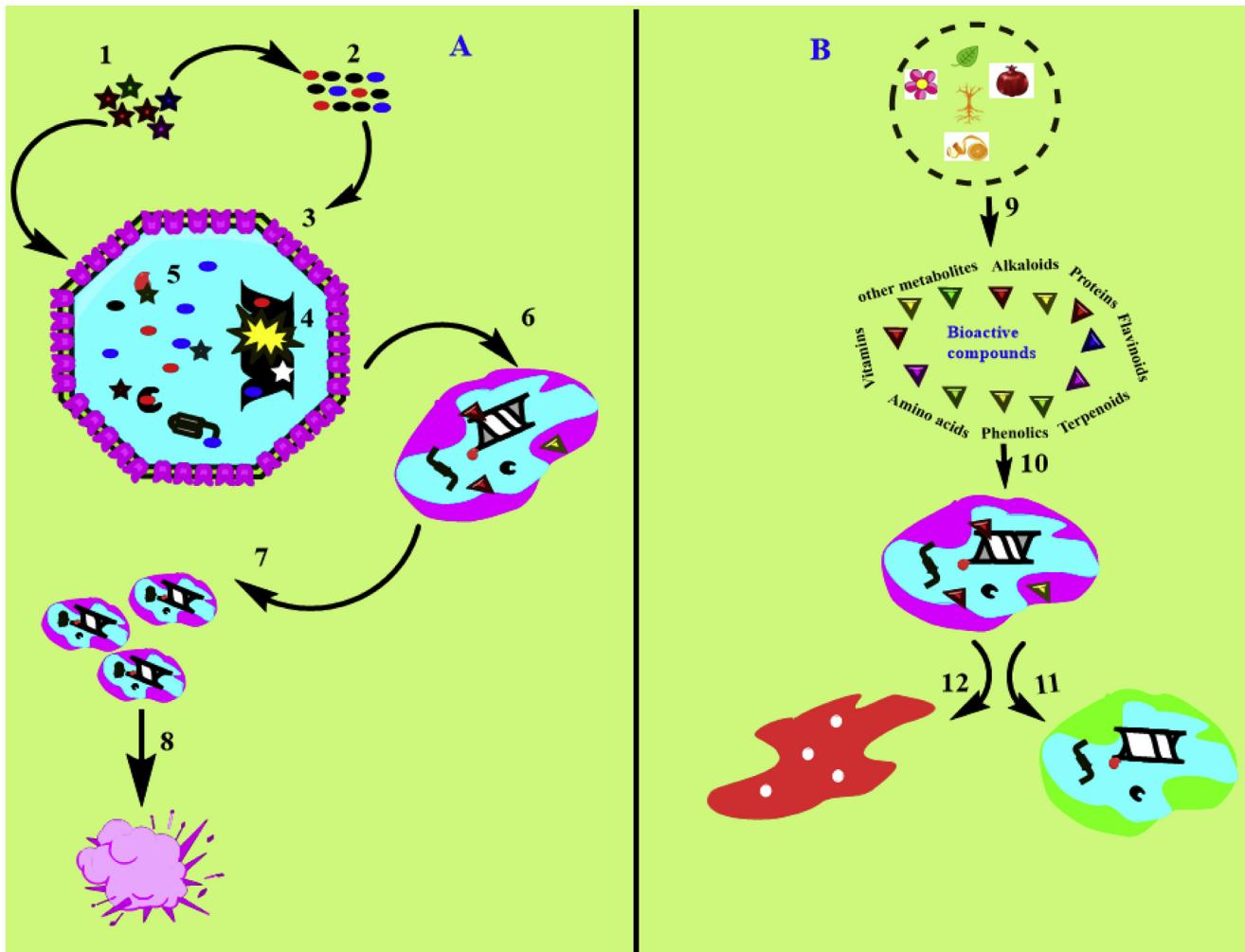


Figure 3. Schematic representation of carcinogenesis model.

(A) In step 1, 2 and 3 various carcinogen and reactive oxygen species ROS entering the cell. Step 4: the entered carcinogen causes genetic mutations that leads to cancer initiation. Step 5: interference of carcinogen and ROS with cellular proteins, enzymes and growth factors. The attack of carcinogen and ROS on DNA, proteins and enzymes of a normal cell. Step 6: transformation of normal cell into cancerous cell. Step 7: Proliferation of cancer cell. Step 8: Tumour cell. (B) Step 9: Different bioactive compounds isolated from different plant materials. Step 10: Application of bioactive compounds on cancerous cell. Step 11: Either cell become normal after phytochemical therapy. Step 12: Or phytochemicals causes apoptosis in cancerous cell.

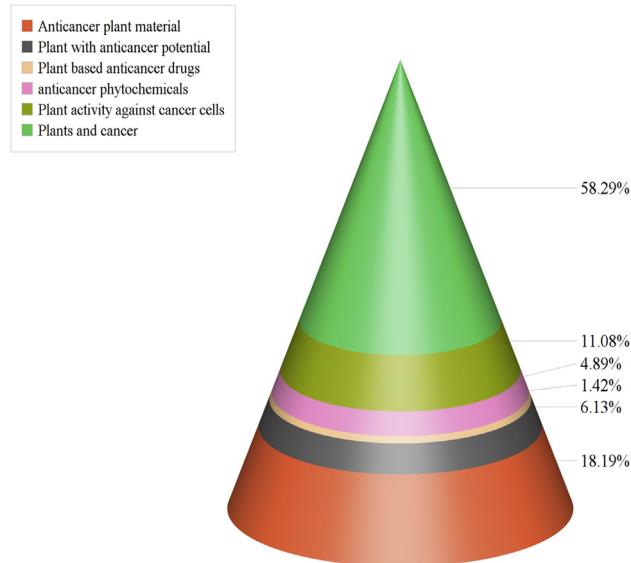


Figure 4. Literature published during 2010–2017 on antineoplastic phytochemicals.

Search results of different keywords used related to plant phytochemicals and cancer (2010–2017).

its antiangiogenic property. Combretastatins (A-4 CA4) is a water soluble analog and is effective against leukemia, lung cancer and colon cancers [97]. The different parts of Indian medicinal plant *Terminalia bellerica* (Combretaceae) fruit, kernel and bark are used in isolation of active combretastatins compounds with strong anticancer properties [16,97]. This agent is presently under research to cure medullary thyroid and aplastic thyroid cancer [178].

3.9. Triterpenoid acids

Triterpenoid acids are also naturally occurring phytomolecules with anticancer properties. Moreover, these potent anti-cancer agents have shown strong anticancer results in both *in-vitro* and *in-vivo* against leukemia, pancreatic and breast cancer. Other anticancer agents like, CDDO (2-cyano-3, 12-dioxoolean1, 9-dien-28-oic acid) and its methyl ester are active against ovarian cancer [113]. Betulinic acid is another triterpenoid isolated from *Ziziphus mauritiana*, *Ziziphus rugosa* and *Ziziphus oenoplia* and *Betula* Sp. (Betulaceae) and is cytotoxic against a wide range of cancer including human melanoma [179].

3.10. Capsaicin

Capsaicin is also a natural phytochemical isolated from red pepper and exert strong anticancer, antimutagenic, anti-metastatic, anti-angiogenic and chemopreventive functions in pancreatic, prostatic, liver, skin, leukemia, lung, bladder, colon, and endothelial cells [180,181]. Capsaicin regulate different molecular targets in breast cancer like, caspase-3, reactive oxygen species (ROS), Rac1, and HER-2 etc. [182]. Capsaicin is more potent by inducing apoptosis in the presence of *p53* gene product (*p53* known as the ‘Grandfather of the genome’ in terms of care) [183]. Capsaicin produced apoptosis in breast cancer (H-Ras, MCF10A cells) by inducing ROS and Rac1 signaling pathways. These ROS and Rac1 pathways are specifically induced by proteins like, p38, c-Jun N-terminal protein kinase-1 [184].

3.11. Flavones/Flavonoids

Flavonoids are also plant-specific secondary metabolites with around 8 000 diverse compounds largely distributed in fruits, grains, tea, vegetables, soybean and play a vital role for the treatment of large number of cancer [110]. Freeze-dried berries rich in anthocyanins have been studied for anti-cancer potential against oral lesions [185], familial adenomatous polyposis [186], and esophageal dysplastic lesions [187]. Flavopiridol is a plant-derived semisynthetic flavone that inhibit cyclin-dependent kinase with anticancer activity against esophageal and gastric cancers [188]. To date, meta-analyses have primarily focused on the functions of dietary flavonoids including inhibition of DNA topoisomerase I, and cyclooxygenase and are used against breast [189], lung [190], stomach and colorectal cancer [191]. Flavopiridol and its mechanism of action on different enzymes are given in Figure 1.

3.12. Cyanidin glycosides

Cyanidin is an organic compound isolated from apples, grapes, plums, blackberry, raspberry, red berries, cranberry, red onion and red cabbage with multiple biological functions as for e.g., antioxidant, radical scavenging effects, inhibit cell growth and division through COX-2 and *iNOS* gene expression in colon cancer cells [192]. Cyanidin-3-orutinoside, cyanidin-3-*O*-glucoside and freeze-dried black raspberries selectively inhibit cell growth and induced apoptosis in a highly tumorigenic rat esophagus RE-149 DHD cell line [192]. Cyanidin glycosides from red berries perform functions of anticancer via many mechanisms. It stops the synthesis of COX-2 enzyme in colon cancer, induces apoptosis in prostate cancer, stops MMP-9 expression in bladder and lung cancer [193]. Stops the Erk phosphorylation and MMP-2 and MMP-9 in fibrosarcoma cells, suppresses expressions of Jun N-terminal kinases (JNK), MMP-9 and Erk enzymes in gastric cancer (stomach cancer) [194]. Cyanidin and its nuclear and cellular targets are summarized in Figure 1.

3.13. Saffron (Crocetin)

Crocus sativus L. commonly known as Saffron, rich in potent anticancer compound carotenoids, crocin, crocetin and safranal [67]. Saffron is marked as a promising agent for a novel anticancer drug against human lung, liver, skin, pancreatic, colorectal and breast cancer by regulating different nuclear and cellular factors, inhibiting iNOS, COX-2 enzymes, reduced

serum level IL- β , TNF- α , cyclin B, cyclinA and cdk2, upregulate Bax/Bcl-2 ratio, regulate of caspase-3, 8 and 9 expression, down-regulate MMP-2, MMP-9 expression, induces apoptosis, targets microtubules and inhibit invasion and metastasis [91,195].

3.14. Epigallocatechin gallate

The main polyphenolic constituent of green tea called epigallocatechin possess the ability to restore genes expression of tumor suppression such as retinoid X receptor alpha, results in breast cancer inhibition by binding to many high affinity target proteins such as, 70 kDa zeta-associated protein (Zap-70) [196]. Molecular docking studies confirmed that both PI3K and mTOR signaling pathways binds well to the PI3K kinase domain active site displaying ATP-competitive activity in MDA-MB-231 and cervical cancer [197], brain cancer [198] and bladder cancer [199].

3.15. Gingerol

Gingerol is also a group of bioactive compound isolated from the fresh rhizome of *Zingiber officinale* containing [6]-gingerol, [8]-gingerol and [10]-gingerol with marked anticancer properties in colon, pancreas, ovarian and breast cancers. It down-regulates the expression of iNOS and TNF-alpha through suppressing NF- κ B nuclear translocation and $I\kappa B\alpha$ phosphorylation [200,201]. Oyagbemi et al. summarized the mechanisms of action of gingerol on K562 cells, MOLT4 cells with high reactive oxygen species, induced apoptosis in leukemia cells by mitochondrial pathway [201]. [10]-gingerol has strong anti-cancer potential than that of [6]-gingerol and [8]-gingerol and have shown promising results for the treatment of MDA-MB-231 and MDA-MB-468 breast cancer. The inhibitory effect of [10]-gingerol on MDA-MB-231 cells was related with the reduction of number of cell divisions, cell cycle arrest, induces apoptosis and releases proapoptotic mitochondrial cytochrome c [202]. Gingerol mechanism of action on different molecular targets is given in Figure 1.

3.16. Lycopene

Lycopene is a bright red pigment with anticancer potential isolated from tomatoes, watermelons, red papayas and red carrots and plays significant role in targeting PI3K/Akt signaling pathway in pancreatic and stomach cancer by down-regulating Erk and Bcl-2 proteins [165]. It up-regulates anti-oxidant enzymes (GSH, GST and GPx) and removes oxidative damage caused by the carcinogens in breast, endometrial, prostate and colon cancer [203]. HT-29 colorectal cancer cells and animal models have shown that lycopene has also effect on cell proliferation and progression by interacting with various cellular signaling pathways like, NF- κ B and JNK [204]. Lycopene also inhibited invasion, metastasis and proliferation in human SW480 colorectal cancer cells by restraining NF- κ B and JNK activation, causes inflammation and suppresses the expression of COX-2, iNOS IL-1 β , IL-6, and TNF- α [205]. Lycopene and its different nuclear and cellular targets are given in Figure 2.

3.17. Vitamin D from mushroom

Mushrooms exposed to light like vertebrates serve a better source for vitamin D after exposure to ultraviolet B light. Light

exposed mushrooms vitamin D has been involved in therapy of wide range of cancer including colon cancer [206], breast cancer [207] pancreatic_ENREF_105 and ovarian cancer by targeting different proteins, enzymes and signaling pathways, prevents carcinogenesis, metastasis and induces apoptosis [208].

3.18. Polysaccharides from mushrooms

Biologically active polysaccharides have been detected in fruiting bodies and mycelial mass of Macromycetes. Mushroom polysaccharides prevent carcinogenesis and display immune cell-mediated anticancer potentials [209]. Polysaccharides with antitumor functions like glucans, lentinan, tegafur, tegafur in combination with lentinan, and schizophyllan have been used for the treatment of lung, breast, and gastric cancers, enhance cellular immunity in the tumor cell, induce apoptosis, prevent invasion and metastasis, act as a macrophage activator, T-cell adjuvant, induce gene expression of cytokines and increase patient's survival with head and neck cancers [210]. Mushrooms polysaccharides and its molecular targets are given in Figure 2.

3.19. Dietary fibers from mushrooms

Mushrooms dietary fibers are also used as strong anticancer agents against different kind of cancers targeting different molecular pathways. Mushroom cell walls contain high molecular weight materials which cannot be digested and absorbed by human intestine, but can absorb carcinogenic substances (heavy metals and free radicals etc.), chitin, homo- and heteropolysaccharides and in this way these high molecular weight materials have been proven as strong anticancer agent against a variety of cancers [211].

3.20. Proteins from mushroom

Mushroom proteins are one of the most extensively studied bioactive substances of mushrooms with their pharmaceutical potential and protein engineering. Many potent anticancer agents such as lectins, bolesatine, hemolysins, phallolysin, nebrodeolysin, laccases, calcaelin and ribonucleases have been isolated from different species of mushrooms (e.g., *Polyporus adusta* and *Ganoderma carpense*, *Pleurotus ostreatus*, *Pleurotus eryngii*, *Pleurotus nebrodensis*, *Amanita phalloides* and *Calvatia caelata* etc.) and are used to treat a variety of cancers including lung cancer [212,213]. *Ganoderma lucidum* extracts can significantly inhibited the release of MMP-2, MMP-9, IL-6 and IL-8 in triple negative breast cancer cells [214]. In the past several chemical compounds extracted from mushroom possessed anticancer properties such as polysaccharopeptide significantly increased the ratio of CD4⁺/CD8⁺/CD14⁺/CD16, T lymphocytes, increase the quantity and percentage of the B lymphocytes and CVP, induce apoptosis and cell cycle arrest [215].

3.21. Vitamin E from plant oil

Vitamin E has been reported as anti-tumor agent and represents a group of compounds consisting of both tocotrienols and tocopherols. It is fat-soluble anti-oxidant present in sunflower oil, germ oil, safflower oils and wheat. It has been researched that both tocopherols and tocotrienols exhibit antitumor properties like proapoptotic, anti-proliferative effects in both either *in-vitro* and *in-vivo* studies [216].

3.22. Fisetin

Fisetin is an active flavone found in various plant species such as strawberries, apple, grape and onion. It has been examined for its potential anticancer consequences; these are anti-migration and anti-proliferation, apoptosis effects on human colon cancer [111,217]. Fisetin is also used to treat human lung cancer by displaying dual inhibition of PI3K/Akt signaling pathways [218]. Moreover, it has anticancer effects in a wide range of cancer cells. For instance, fisetin found to induce apoptosis through inhibition of MAPK signaling network in human lung cancer and reactive oxygen species production in human oral cancer. It also induces apoptosis in human renal carcinoma caki cells via p53-mediated up-regulation of DR5 expression [219]. The mechanisms of action of fisetin on different signaling pathways are given in Figure 1.

3.23. Resveratrol

Resveratrol is also a naturally occurring polyphenol and has been identified in mulberries, peanuts, grapes, bilberries and blueberries. Resveratrol play substantial role in curing a wide range of cancers including breast, colorectal, liver, pancreatic, prostate cancer and lung carcinoma by up-regulating p53 and Bcl-2 associated X proteins and down-regulating MMPs, NF-κB, AP-1, Bcl-2, cyclins, cyclin dependent kinases, cytokines, and COX-2 proteins [220,221]. Resveratrol is known to inhibit angiogenesis, suppressing VEGF protein action by reducing MAP kinase phosphorylation [222]. The mechanism of action of resveratrol on different nuclear and cellular factors are given in Figure 1.

3.24. Anticancer compounds from algae

More than 50% of the marine blue-green algae are largely used for the isolation of anticancer compounds which are effective in either inducing apoptosis or triggering signaling pathways through activation of protein kinase-c enzymes, NF-κB, MAPK kinases, p53, cytokines release and ROS production. The cell extracts of *Calothrix* [Calothrixin A (I) and B (II), *Lynbya majuscule* (Curacin-A)] are strong antiproliferative agents with inhibitory role in colon, renal and breast cancers [97]. Similarly, cryptophycin 1 isolated from a species of *Nostoc* (GSV 224) revealed strong cytotoxicity against human solid tumors. The compounds isolated from edible seaweed like *Padina boergesenii*, *Ulva reticulata*, *Gracilaria foliifera*, *Palmaria palmate*, *Acanthophora spicifera*, *Sargassum thunbergii*, *Ascophyllum nodosum* and *Ectonia cava* have also shown marked anticancer activities and are used to treat kidney cancer, ammary adenocarcinoma, colon adenocarcinoma, human nasopharyngeal and colorectal cancer [223,224]. Cyanobacterium (e.g. *Spirulina platensis*) is effective against different types of human cancers (liver, lung, stomach and breast cancer via the production of valuable products (phycobiliproteins including c-phycocyanin, phycocyanobilin, allophycocyanin) [225].

3.25. Apigenin

Apigenin (APG) is a naturally occurring flavonoid identified in the different fruits and vegetables such as celery, chamomile and parsley with features including low toxicity and non-mutagenic, induces apoptosis and targets leptin/leptin receptor pathway in

lung cancer. Apigenin also activates caspase dependent extrinsic apoptosis pathway, inhibit signal transducer and activator of transcription 3 (STAT3) signaling pathways [226,227]. APG is also used to treat MDA-MB-453 breast cancer through inhibiting STAT3 signaling pathway by expression levels of caspase-3, caspase-8, induces extrinsic apoptosis, blocking the phosphorylation of JAK2 and STAT3 pathways [226]. Detailed information about apigenin regulating different protein, enzymes and signaling pathway are given in Figure 1 and 2.

3.26. Curcumin

Curcumin is also a lead phytochemical extracted from *Curcuma longa* with inhibitory property over the growth of human glioblastoma cells by modulating several nuclear and cellular factors, upregulates the expressions of different genes and their products [*p16*, *p21* and *p53* (These genes are also called as ‘Grandfather of the genome’ in terms of care for the cell], *Bcl-2* associated X protein (Bax protein), EIK-1 (ETS oncogenic family), extracellular signal regulated kinase (Erk enzyme), early growth response protein 1, c-Jun N-terminal kinase, and caspase enzymes (Caspase-3, 8, 9) and decreases the level of *Bcl-2*, mTOR, p65, protein kinase B (Akt), retinoblastoma protein (pRB), NF-κB, and cyclin D1 proteins [54]. Detailed information about curcumin and its mechanism of action on different proteins and signaling cascades are summarized in Figures 1 and 2.

3.27. β-elemene

β-elemene a sesquiterpene, is also a promising anticancer agent with a wide range of its effect against drug-resistant tumors and has been isolated from *Curcuma wenyujin* [228]. β-elemene is major component of traditional Chinese medicine and inhibit different forms of cancer, induces apoptosis and cell death, inhibit the expression of VEGF, downregulates Akt phosphorylation and CD34 expression, suppressing PI3K/Akt/mTOR, MAPK and pathway, attenuating angiogenesis and upregulates the E3 ubiquitin ligases, Cbl-b and c-Cbl in human gastric cancer [229,230]. β-elemene molecular targets are given in Figure 2.

3.28. Chalcone

Chalcone is also a naturally occurring anticancer flavonoid in fruits and vegetables. It is responsible for activation of different caspases (caspase-8, 9, 12 enzymes), upregulate the of pro-apoptotic proteins expression (Bid, Bax, and Bak proteins), decreases anti-apoptotic *Bcl-2* gene expression and have been used for the treatment of the treatment of colon, lung, breast, liver and prostate cancer [231]. Chalcone targeted different nuclear and cellular factors (Bax, Bid, and Bak, *Bcl-2* proteins, caspase-8, 9, 12 enzymes are illustrated in Figure 2.

3.29. Sesquiterpene lactones

Sesquiterpene lactones constitute large and diverse group of bioactive compounds isolated from several plant families (*e.g.* Asteraceae) exhibiting cancer cell cytotoxicity and antineoplastic efficacy and are used for the treatment of a large number of cancer like, prostate, liver, lung, breast esophageal cancer [232].

Sesquiterpene lactone *e.g.* deoxyelephantopin and isodeoxyelephantopin are components of *Elephantopus carolinianus* and *Elephantopus scaber* and have been shown to induce apoptosis via multiple mechanisms, comprised induces ROS, mitochondrial dysfunction, modulate Bcl-2 family protein, arrest cell cycle, inhibit NF-κB, and STAT3 activation [233].

3.30. Chrysin

Chrysin [5,7-dihydroxyflavone], is an effective anticancer compound showed less side effects and distributed in propolis, honey and blue passion flower, chamomile and possess strong antitumor effects on different cancer cell lines (DU145 and PC-3) [234,235]. This flavone has also induced apoptosis in SW480 colorectal cancer, arrest cell cycle at G₂/M phase, results in DNA cleavage and apoptosis, increasing ROS production and lipid peroxidation, suppressed the abundance of S6, AKT, PI3K, P90RSK and P70S6K, proteins, stimulate MAPK and ERK1/2 and P38 proteins in the prostate cancer cells [161,236]. Chrysin and its mechanism of action on different targets are given in Figure 1.

3.31. Scutellarin glycoside

Scutellarin is a promising anticancer agent isolated from medicinal plant species, *Scutellaria barbata* and *Scutellaria altissima* and exhibits anti-tumor functions on different cancers for example, human colon cancer, liver cancer, and prostate cancer [237]. Scutellarin suppressed cancer cell proliferation, induces cell cycle arrest at G₂/M phase, upregulating caspase-3, 9, and Bax/Bcl-2 ratio in prostate cancer [238]. Scutellarin also induced apoptosis in liver carcinoma (HepG2 cells) via STAT3 signaling cascade and caspase-3 enzyme activation [177]. In addition, this chemical agent also induces apoptosis in human colorectal cancer, inhibiting cell growth and induces apoptosis by regulating the *p53* gene product [239]. The molecular targets of scutellarin are given in Figure 2.

3.32. Oroxylum flavone

Oroxylum A is a potential flavone isolated from the *Scutellariae radix* down-regulate the expression of COX-2 and iNOS genes, block NF-κB, inhibit the activation of LPS-induced NF-κB by blocking IκB degradation [240]. Oroxylum A in combination with 5-FU is also used to treat colorectal cancer, exhibiting double action with COX-2 inhibition and increased ROS generation. Thus, oroxylum combination therapy could be a promising tool in order to reduce 5-FU doses and subsequent *in-vivo* side effects [241]. Oroxylum and its molecular targets are illustrated in Figure 2.

3.33. Kaempferol

Kaempferol is also a naturally occurring anticancer agent isolated from propolis, black tea, grapefruit, broccoli. Kaempferol possess significant antitumor potential on a large number of cancer cells *e.g.* colorectal cancer and HT-29 cancer cells by activating the expression of caspase-3 enzyme, *p53* gene and its products [242,243], arrest cell cycle at G₁ and G₂/M phase by inhibiting the activity of different enzymes (CDK2, Cdc2 and CDK4) [244]. Kaempferol and its nuclear and cellular targets are illustrated in Figure 1.

3.34. Genistein

Genistein is potent antitumor agent isolated from soybeans, lentils, beans and chickpeas. This isoflavone possessed pro-apoptotic function against colorectal cancer [245]. Genistein perform numerous functions as for example, up-regulate the expression of pro-apoptotic proteins (Bax and p21), inhibiting NF- κ B and topoisomerase II enzymes [246,247], upregulate antioxidant enzyme expression such as glutathione peroxidase [248]. Genistein and its action on different protein, enzyme and signaling pathways are given in Figures 1 and 2.

3.35. Silymarin

Silymarin is also naturally occurring flavolignan extracted from *Silybum marianum*. This flavolignans mixture contained silydianin, silychristin, silibinin (silybin A and B), and iso-silybin (A and B), [249]. Silymarin induces cell cycle arrest and apoptosis by acting on cyclin dependent kinases and has been used together with paclitaxel and doxorubicin to treat colorectal cancer [250].

3.36. Ursolic acid

The ursolic acid is triterpene is the main constituent in herbal species likewise rosemary and basil plants. This antioxidant compound play significant role in the modulation of cellular redox status of normal cells and exerts pro-oxidative action on tumor cells. It exhibits pro-apoptotic effects on colorectal cancer (HCT116 cell line) by reducing the level of pro-inflammatory NF- κ B cytokine, survival effectors Bcl-2 and pro-metastatic MMP-9 matrix metalloprotease [251]. Ursolic acid and molecular mechanism of action on different nuclear and cellular targets are given in Figure 1.

3.37. Ginsenosides

Ginsenosides constitute a class of active compounds responsible for pharmacological activities obtained from ginseng root. Pre-clinical and clinical researches demonstrated that ginsenosides have cancer preventing activities to various tumors, including liver, breast, gastric, ovarian, melanoma and colon cancer [252]. The six major ginsenosides (Rb1, Rb2, Rc, Rd, Re, and Rg1) constitute around 80% of the total ginsenosides in ginseng root and various minor ginsenosides [Rg3(S), Rh2(S), F2, compound K (C-K), Rg2(S), Rh1(S), F1, protopanaxatriol and gypenoside XVII] possess anti-invasion and anti-migration properties, induces apoptosis and cell cycle arrest [253]. Panaxadiol is a strong anticancer agent isolated from *Panax ginseng* and *Panax pseudoginseng* and has potent anti-cancer activity in different cancer cell lines and signaling pathways [254]. The same results were found with protopanaxadiol metabolite that significantly enhanced 5-FU effects on HCT116 cells by arresting cell cycle at G₁ phase and induces apoptosis [60]. One other member ginsenoside Rg3, is able to block NF- κ B expression in HCT116 cell line leading to apoptosis [255]. Furthermore, ginsenoside-Rg5 is used to treat human cervical cancer by inducing apoptosis and cause DNA damage [256].

3.38. Celastrol

Celastrol is also a strong anticancer compound in terms of its efficacy isolated from the bark of *Tripterygium wilfordii* and

inhibits heat shock protein, blocking its interaction with Cdc37, apoptosis induction via caspase-3 enzyme in ovary (OVCAR-8), colon (SW620), lung 95-D and prostate cancer [257]. Celastrol also induces apoptosis and inhibit the expression of onco-protein in acute myelogenous leukemia1-ETO/C-KIT [8,21] [258]. It is also used to treat lung cancer cells through Hsp90 client protein degradation and caspases-dependent pathways [259]. Data from different cancers reports such as breast, lung, colon, prostate, esopharyngeal, glioblastoma, liver, skin, myeloma, pancreas, liver, leukemia, and gastric cancer) and animal models have suggested that celastrol induces apoptosis and cell cycle arrest, autophagy by the activation of ROS/c-JNK signaling pathway, inhibit angiogenesis and exhibit anti-invasive effect, upregulate death receptors in breast and colon cancer, activate Fas/Fas ligand pathway in lung cancer and inhibit PI3K/Akt/mTOR pathway in triple negative breast cancer [260].

3.39. Gossypol

Gossypol is also a natural phytochemical found in cotton seeds (*Gossypium*) and *Thespesia populnea*, displays potential anti-cancer activities, has completed phase II clinical trials for treatment of human breast and prostate cancer. Its antitumor properties have been studied in a variety of tumors (lymphoid, hematologic and solid tumors). Gossypol suppresses cell proliferation, induces autophagy and apoptosis in colorectal cancer, HT-29, HCT116 and RKO cancer cell lines [61].

3.40. Polysaccharides from plants

Plant polysaccharides are also promising anticancer agents and have long been used as therapy for variety of cancers e.g. liver (HepG2 cells), lung (A549) cells, HL-60 cells, H157 cells, ovarian cancer, and human lymphatic endothelial cells [58]. Polysaccharides as anticancers agents haven been reported variously from medicinal plants [261]. Recently, researchers are shifting their research direction from microbe's polysaccharides to plant polysaccharides as they are non-toxic [262]. Many *in-vitro* and *in-vivo* studies indicated that these polysaccharides inhibit tumor cell proliferation by inducing cell cycle arrest and apoptosis in different type of cancers [66,263]. There are numerous reports on the role of polysaccharides in inducing cell cycle arrest and apoptosis, regulate different signaling cascades and cell cycle genes and inhibit cancer cell proliferation in a wide range of cancer cell lines (HepG2 cells, A549 cells, human lymphatic endothelial cells and ovarian cancer [239,264–266].

3.41. Isothiocyanates

Isothiocyanates is a potent phytochemical occurs in vegetables belongs to family Cruciferae such as watercress and broccoli and are used for the treatment of different cancer namely, colorectal cancer, cervical cancer, lung cancer, prostate cancer, and human T-Leukemia cells without causing any toxic side effects [267]. These natural isothiocyanates can induce apoptosis and ROS-mediated mechanism, arresting cell cycle at G₂/M phase, downregulate activated signaling cascades and modulate epigenetic changes, inhibit cell proliferation, progression and invasion-metastasis in colorectal and prostate cancer [268]. Sulforaphane, a

dietary isothiocyanate possess anticancer property in cervical cancer cells via inducing G₂/M arrest, downregulate cyclinB1 and up-regulate GADD45 β proteins [269].

3.42. Genipin

Genipin is a natural phytochemical isolated from *Gardenia jasminoides* and is used to treat breast cancer [270]. In breast cancer, genipin regulates different protein and enzymes as for examples, caspase-3, Bax, Bcl-2, JNK, p38, MAPK. Genipin has anti-proliferative activity in MDA-MB-231 breast cancer cells by down-regulating Bcl-2 expression and up-regulating Bax and caspase-3, pro-apoptotic signaling cascades such as JNK and p38 MAPK [271].

3.43. Denbinobin

Denbinobin is another multifunctional phytochemical isolated from the stem of *Ephemerantha lonchophylla* and *Dendrobium moniliforme* with potent anti-cancer, anti-angiogenesis and apoptosis-inducing properties [272]. This compound inhibits metastasis by inhibiting Src kinase activity, decreases iNOS and COX-2 activity in concentration-dependent manner by suppressing NF- κ B activation in human breast cancer cells [273].

4. Different strategies for the development of anticancer phytochemicals

The potential of medicinal plants as therapeutic agents depends upon the quality and quantity of active phytochemicals in them, which vary with latitude, longitude, altitude, age, climate and season from species to species. Pharmacological functions and their level vary with plant parts. These bioactive phytomolecules can also be used in anticancer therapeutics but they still demand further research. The purification of active phytomolecules may involve various strategies such as combinatorial chemistry, isolation assays, and bioassay-guided fractionation. Bioassay guided fractionation with various analytical techniques could be used to separate various bioactive compounds from the mixture of compounds. The process begins with the natural extracts test (from dry/wet plant material) with confirmed biological activity. Then, suitable matrices are used for the fractionation of active extracts, tested for bioactivity and various analytical techniques such as TLC, HPLC, FTIR, Mass spectroscopy and NMR etc. must be used for the separation of active fractions. For polarity order rise different solvents should to use. Superdex, Sephadex, Silica or any other suitable matrix can be used for fractionation. There are so many dyeing agents used for the detection of natural compounds in medicinal plants e.g. Vanillinesulfuric acid. These procedures

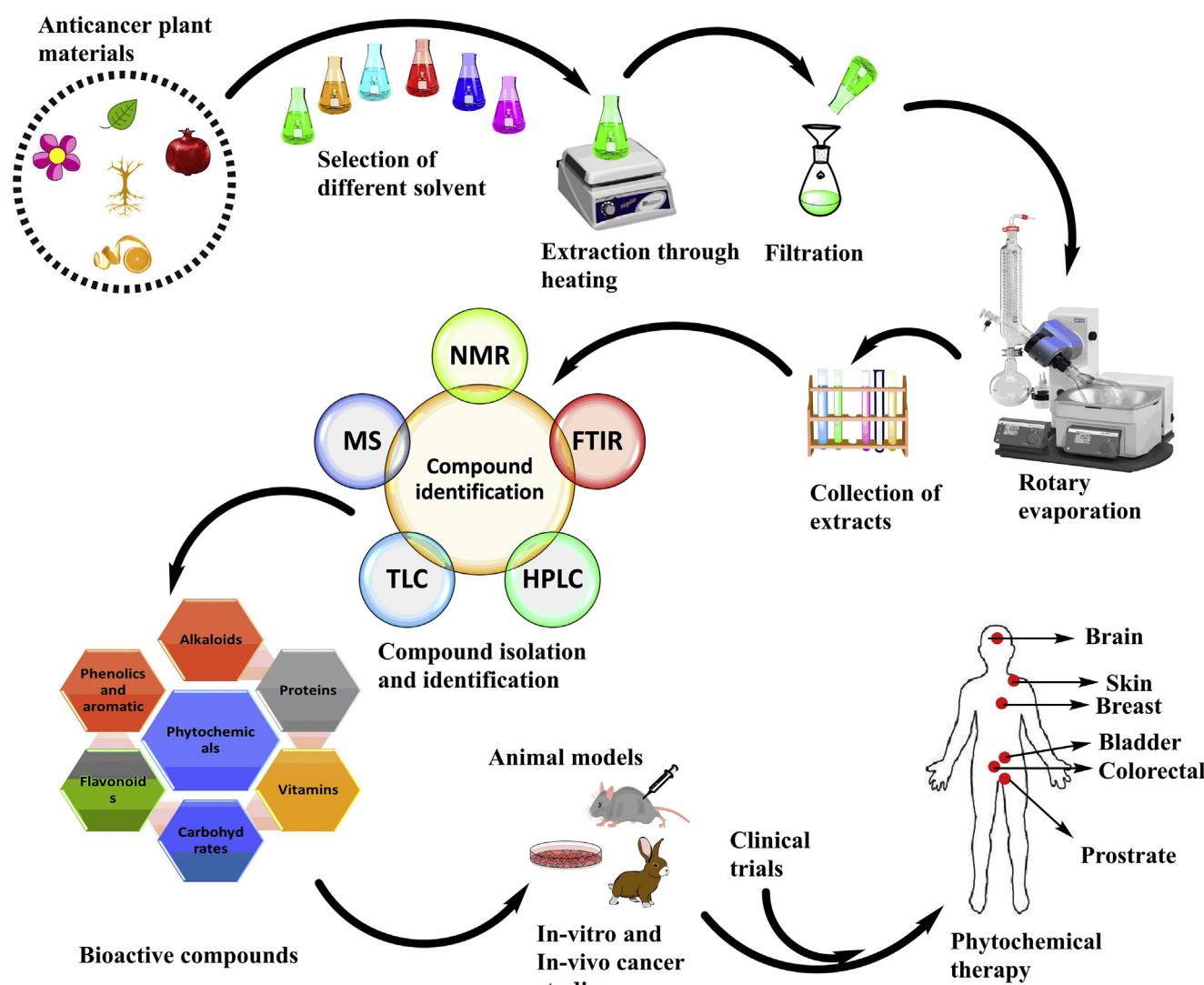


Figure 5. Detailed scheme of anticancer phytochemical synthesis, optimization, characterization and prospective use as cancer therapeutic agent.

could be change however purity, quality and quantity of the bioactive compounds should be high as much as possible and this can be achieved by using high quality of solvents, matrices and careful handling. After purification of these phytomolecules they must be examined for *in-vitro* or *in-vivo* anticancer effects. If a better anticancer property is achieved by the molecule then other aspects like pharmacokinetics, pharmacodynamics, immunogenicity, metabolic fate, biosafety and side effects, drug interactions, dose concentration etc. must be researched for future drug designing. Detailed scheme of bioactive compound synthesis, optimization, characterization, testing, and potential application as a cancer therapeutic agent is shown in Figure 5.

5. Conclusions and future prospects

It has been evident from the present review that phytochemicals serve as promising and effective research area with bright future. The growing incidence of cancer and high cost, various limitations in the conventional therapy including high cost, and high toxicity of present anticancer drugs has faced a severe challenge to all the researchers to design and develop an alternative, eco-friendly, biocompatible and cost-effective strategy in a greener way. Under this scenario, phytomolecules are expected to revolutionize cancer treatment in the next decade. High biodegradability and biocompatibility have increased the efficacy of these phytomolecules in cancer therapy. This comprehensive review paper provides information on medicinal plants and their bioactive compounds with potential to cure different types of cancer. Potential anticancer phytochemicals described in this comprehensive review article should be further researched in clinical trials (Curcumin, epigallocatechin, isothiocyanates, gossypol, sulforaphane, garcinol, etc.) on different models for their effectiveness and toxicological documentation. Furthermore, extensive research work should be carried out on these phytochemicals to evaluate their possible applications, toxicological and particular genotoxic profile against a wide range of cancer in both either *in-vitro* or *in-vivo*.

Conflict of interest statement

The authors declare no competing interest.

Acknowledgements

All authors listed have made substantial, direct and intellectual contribution to the work. Javed Iqbal and Banzeer Ahsan Abbasi summarized the literature, wrote the manuscript and drew the figures. Sobia Kanwal, Barkat Ali and Ali Talha Khalil revised the manuscript. TM helped in interpretation by reviewing several draft of the manuscript.

Appendix A. Supplementary data

Supplementary data related to this article can be found at <https://doi.org/10.1016/j.apjtb.2017.10.016>.

References

- [1] He L, Gu J, Lim LY, Yuan ZX, Mo J. Nanomedicine-mediated therapies to target breast cancer stem cells. *Front Pharmacol* 2016; **7**: 313.
- [2] Qin W, Huang G, Chen Z, Zhang Y. Nanomaterials in targeting cancer stem cells for cancer therapy. *Front Pharmacol* 2017; **8**: 1.
- [3] Zhang LQ, Lv RW, Qu XD, Chen XJ, Lu HS, Wang Y, et al. Aloesin suppresses cell growth and metastasis in ovarian cancer SKOV3 cells through the inhibition of the MAPK signaling pathway. *Anal Cell Pathol* 2017; **2017**: 1-6.
- [4] American Cancer Society. *Cancer facts & figures 2016*. Atlanta, GA: American Cancer Society; 2016.
- [5] Siegel RL, Miller KD, Jemal A. Cancer statistics, 2016. *CA Cancer J Clin* 2016; **66**(1): 7-30.
- [6] Siegel RL, Miller KD, Fedewa SA, Ahnen DJ, Meester RG, Barzi A, et al. Colorectal cancer statistics, 2017. *CA Cancer J Clin* 2017; **67**(3): 177-93.
- [7] Krishnamurthi K. 17-screening of natural products for anticancer and antidiabetic properties. *Cancer* 2007; **3**: 4.
- [8] Horn L, Pao W, Johnson DH. Neoplasms of the lung. Chapter89. In: Longo DL, Kasper DL, Jamson JL, Fauci AS, Hauser SL, Loscalzo J, editors. *Harrison's principles of internal medicine*. 18th ed. New York, NY: MacGraw-Hill; 2012.
- [9] Zhou Z, Tang M, Liu Y, Zhang Z, Lu R, Lu J. Apigenin inhibits cell proliferation, migration, and invasion by targeting Akt in the A549 human lung cancer cell line. *Anti Cancer Drugs* 2017; **28**(4): 446-56.
- [10] Kharb M, Jat RK, Gupta R. A review on medicinal plants used as a source of anticancer agents. *Int J Drug Res Technol* 2012; **2**: 177-83.
- [11] Dixit S, Ali H. Anticancer activity of medicinal plant extract-a review. *J Chem Chem Sci* 2010; **1**: 79-85.
- [12] Parkin D, Bray MF, Ferlay J, Pisani P. Global cancer statistics. *CA Cancer J Clin* 2015; **55**: 74-108.
- [13] Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer* 2015; **136**: E359-86.
- [14] Faria SS, Morris CFM, Silva AR, Fonseca MP, Forget P, Castro MS, et al. A timely shift from shotgun to targeted proteomics and how it can be groundbreaking for cancer research. *Front Oncol* 2017; **7**: 13.
- [15] Thakore P, Mani RK, Kavitha SJ. A brief review of plants having anti-cancer property. *Int J Pharm Res Dev* 2012; **3**: 129-36.
- [16] Cragg GM, Newman DJ, Yang SS. Natural product extracts of plant and marine origin having antileukemia potential. The NCI experience. *J Nat Prod* 2016; **69**: 488-98.
- [17] Khan H. Medicinal plants in light of history: recognized therapeutic modality. *J Evid Based Complement Altern Med* 2014; **19**: 216-9.
- [18] Weaver BA. How taxol/paclitaxel kills cancer cells. *Mol Biol Cell* 2014; **25**(18): 2677-81.
- [19] Singh S, Bhupender S, Kanwar SS, Kumar A. Lead phytochemicals for anticancer drug development. *Front Plant Sci* 2016; **7**: 1667.
- [20] Vinogradov S, Wei X. Cancer stem cells and drug resistance: the potential of nanomedicine. *Nanomedicine* 2012; **7**: 597-615.
- [21] Patra CR, Mukherjee S, Kotcherlakota R. Biosynthesized silver nanoparticles: a step forward for cancer theranostics? *Nanomedicine* 2014; **9**: 1445-8.
- [22] Mukherjee S, Patra CR. Therapeutic application of anti-angiogenic nanomaterials in cancers. *Nanoscale* 2016; **8**: 12444-70.
- [23] Caruso M, Colombo AL, Fedeli L, Pavesi A, Quaroni S, Saracchi M, et al. Isolation of endophytic fungi and actinomycetes taxane producers. *Ann Microbiol* 2000; **50**: 3-14.
- [24] Aung TN, Qu Z, Kortschak RD, Adelson DL. Understanding the effectiveness of natural compound mixtures in cancer through their molecular mode of action. *Int J Mol Sci* 2017; **18**(3): 656.
- [25] Tariq A, Sadia S, Pan K, Ullah I, Mussarat S, Sun F, et al. A systematic review on ethnomedicines of anti-cancer plants. *Phytother Res* 2017; **31**: 202-64.
- [26] Ayoob I, Hazari YM, Lone SH, Khuroo MA, Fazili KM, Bhat KA, et al. Phytochemical and cytotoxic evaluation of peganum harmala: structure activity relationship studies of harmine. *Chem Sel* 2017; **2**(10): 2965-8.
- [27] Oko E, Kadioglu O, Greten HJ, Efferth T. Pharmacogenomic characterization and isobologram analysis of the combination of ascorbic acid and curcumin-two main metabolites of *Curcuma longa*-in cancer cells. *Front Pharmacol* 2017. <https://doi.org/10.3389/fphar.2017.00038>.

- [28] Bhandari J, Muhammad B, Thapa P, Shrestha BG. Study of phytochemical, anti-microbial, anti-oxidant, and anti-cancer properties of *Allium wallichii*. *BMC Compl Altern Med* 2017; **17**(1): 102.
- [29] Efferth T. From ancient herb to versatile, modern drug: *Artemisia annua* and artemisinin for cancer therapy. *Semin Canc Biol* 2017. <https://doi.org/10.1016/j.semcaner.2017.02.009>.
- [30] Kumari M, Pattnaik B, Rajan SY, Shrikant S, Surendra SU. *EGCG-A Promis anti-cancer Phytochem* 2017; **3**(2): 8-10.
- [31] Preethi R, Padma PR. Biosynthesis and bioactivity of silver nanobioconjugates from grape (*vitis vinifera*) seeds and its active component resveratrol. *Int J Pharma Sci Res* 2016; **7**(10): 4253.
- [32] Xiong M, Wang L, Yu HL, Han H, Mao D, Chen J, et al. Ginkgetin exerts growth inhibitory and apoptotic effects on osteosarcoma cells through inhibition of STAT3 and activation of caspase-3/9. *Oncolo Rep* 2016; **35**(2): 1034-40.
- [33] Wu F, Zhou L, Jin W, Yang W, Wang Y, Yan B, et al. Anti-proliferative and apoptosis-inducing effect of theabrownin against non-small cell lung adenocarcinoma A549 cells. *Front Pharmacol* 2016; **7**: 465.
- [34] Beg MA, Teotia UV, Farooq S. *In vitro* antibacterial and anti-cancer activity of *Ziziphus*. *J Med Plants* 2016; **4**(5): 230-3.
- [35] Al Sinani SS, Eltayeb EA, Coomber BL, Adham SA. Solamargine triggers cellular necrosis selectively in different types of human melanoma cancer cells through extrinsic lysosomal mitochondrial death pathway. *Cancer Cell Inte* 2016; **16**(1): 11.
- [36] Mehdad A, Brumana G, Souza AA, Barbosa JA, Ventura MM, de Freitas SM, et al. A Bowman-Birk inhibitor induces apoptosis in human breast adenocarcinoma through mitochondrial impairment and oxidative damage following proteasome 20S inhibition. *Cell Death Dis* 2016; **2**: 15067.
- [37] Jaradat NA, Al-Ramahi R, Zaid AN, Ayesh OI, Eid AM. Ethnopharmacological survey of herbal remedies used for treatment of various types of cancer and their methods of preparations in the West Bank-Palestine. *BMC Comp Altern Med* 2016; **16**(1): 93.
- [38] Zhang YY, Huang CT, Liu SM, Wang B, Guo J, Bai JQ, et al. Licocalcone A exerts antitumor activity in bladder cancer cell lines and mice models. *Trop J Pharm Res* 2016; **15**(6): 1151-7.
- [39] Yong YL, Tan LT, Ming LC, Chan KG, Lee LH, Goh BH, et al. The effectiveness and safety of topical capsaicin in posttherapeutic neuralgia: a systematic review and meta-analysis. *Front Pharm* 2017. <https://doi.org/10.3389/fphar.2016.00538>.
- [40] Srikanth S, Chen Z. Plant protease inhibitors in therapeutics-focus on cancer therapy. *Front Pharma* 2016. <https://doi.org/10.3389/fphar.2016.00470>.
- [41] Pahari P, Saikia UP, Das TP, Damodaran C, Rohr J. Synthesis of psoralidin derivatives and their anticancer activity: first synthesis of lespeflorin I 1. *Tetrahedron* 2016; **72**(23): 3324-34.
- [42] Amin S, Barkatullah HK. Pharmacology of *Xanthium* species. A review. *J Phytopharm* 2016; **5**: 126-7.
- [43] Thangapazham RL, Sharad S, Maheshwari RK. Phytochemicals in wound healing. *Adv Wound Care* 2016; **5**(5): 230-41.
- [44] Tu LY, Pi J, Jin H, Cai JY, Deng SP. Synthesis, characterization and anticancer activity of kaempferol-zinc (II) complex. *Bioorg Med Chem Lett* 2016; **26**(11): 2730-4.
- [45] Lee IC, Choi BY. WithaferinA-a natural anticancer agent with pleiotropic mechanisms of action. *Int J Mol Sci* 2016; **17**(3): 290.
- [46] Wal A, Srivastava RS, Wal P, Rai A, Sharma S. Lupeol as a magic drug. *Pharm Biol Eval* 2015; **2**: 142-51.
- [47] Rastogi N, Duggal S, Singh SK, Porwal K, Srivastava VK, Maurya R, et al. Proteosome inhibition mediates p53 reactivation and anticancer activity of 6-gingerol in cervical cancer cells. *Oncotarget* 2015; **6**: 43310-25.
- [48] Tsai CC, Chuang TW, Chen LJ, Niu HS, Chung KM, Cheng JT, et al. Increase in apoptosis by combination of metformin with silibinin in human colorectal cancer cells. *World J Gastroenterol* 2015; **21**: 4169-77.
- [49] Osman NHA, Said UZ, El-Waseef AM, Ahmed ESA. Luteolin supplementation adjacent to aspirin treatment reduced dimethyl hydrazine-induced experimental colon carcinogenesis in rats. *Tumour Biol* 2015; **36**: 1179-90.
- [50] Lin X, Peng Z, Su C. Potential anticancer activities and mechanisms of costunolide and dehydrocostuslactone. *Int J Mol Sci* 2015; **16**: 10888-906.
- [51] Mukhija M, Singh MP, Dhar KL, Kalia AN. Cytotoxic and antioxidant activity of *Zanthoxylum alatum* stem bark and its flavonoid constituents. *J Pharm Phytochem* 2015; **4**: 86.
- [52] Garg P, Deep A. Anti-cancer potential of boswellic acid: a mini review. *Hygeia J D Med* 2015; **7**(2): 18-27.
- [53] Ramasamy K, Agarwal R. Multitargeted therapy of cancer by silymarin. *Cancer Lett* 2008; **269**(2): 352-62.
- [54] Vallianou NG, Evangelopoulos A, Schizas N, Kazazis C. Potential anticancer properties and mechanisms of action of curcumin. *Anticancer Res* 2015; **35**: 645-51.
- [55] Monika Singh J. Plants and phytochemicals as potential source of anticancer agents. *Inte J Adv Res* 2015; **4**: 307-17.
- [56] Choi JY, Hong WG, Cho JH, Kim EM, Kim J, Jung CH, et al. Podophyllotoxin acetate triggers anticancer effects against non-small cell lung cancer cells by promoting cell death via cell cycle arrest, ER stress and autophagy. *Inter J Oncol* 2015; **47**: 1257-65.
- [57] Hoshyar R, Mollaei H. A comprehensive review on anticancer mechanisms of the main carotenoid of saffron, crocin. *J Pharm Pharmacol* 2017; **1**: 1-9.
- [58] Liu YQ, Tian J, Qian K, Zhao XB, Susan LM, Yang L. Recent progress on c-4 modified Podophyllotoxin analogs as potent antitumor agents. *Med Res Rev* 2015; **35**: 1-62.
- [59] Król SK, Kielbus M, Rivero-Müller A, Stepulak A. Comprehensive review on betulin as a potent anticancer agent. *Bio Med Res Int* 2015; **2015**: 584189. <https://doi.org/10.1155/2015/584189>.
- [60] Wang C, Zhang Z, Wan Z, Zhang JY, Anderson CF, He X, et al. Protopanaxadiol, an active ginseng metabolite, significantly enhances the effects of fluorouracil on colon cancer. *Nutrients* 2015; **7**: 799-814.
- [61] Lan L, Appelman C, Smith AR, Yu J, Larsen S, Marquez RT, et al. Natural product 1064 (-)-gossypol inhibits colon cancer cell growth by targeting RNA-binding protein Musashi-1. *Mol Oncol* 2015; **9**: 1406-20.
- [62] Leon IE, Cadavid-Vargas JF, Tiscornia I, Porro V, Castelli S, Katkar P, et al. Oxidovanadium(IV) complexes with chrysin and silibinin: anticancer activity and mechanisms of action in a human colon adenocarcinoma model. *J Biol Inorg Chem* 2015; **20**: 1175-91.
- [63] Yan CH, Li F, Ma YC. Plumbagin shows anticancer activity in human osteosarcoma (MG-63) via the inhibition of s-phase checkpoints and downregulation of c-myc. *Int J Clin Exp Med* 2015; **8**: 14432-9.
- [64] Ghasemzadeh A, Jaafar HZE, Rahmat A. Optimization protocol for the extraction of 6-gingerol and 6-shogaol from *Zingiber officinale* var. rubrum and improving antioxidant and anticancer activity using response surface methodology. *BMC Complement Altern Med* 2015; **15**: 258.
- [65] Perrone D, Ardito F, Giannatempo G, Dioguardi M, Troiano G, Russo LL, et al. Biological and therapeutic activities and anti-cancer properties of curcumin. *Exp Ther Med* 2015; **10**: 1615-23.
- [66] Wozniak L, Skapska S, Marszalek K. Ursolic acid-a pentacyclic triterpenoid with a wide spectrum of pharmacological activities. *Molecules* 2015; **20**: 20614-41.
- [67] Pang SQ, Wang GQ, Lin JS, Diao Y, Xu RA. Cytotoxic activity of the alkaloids from *Broussonetia papyrifera* fruits. *Pharm Biol* 2014; **52**: 1315-9.
- [68] Jung SK, Lee MH, Kim JE, Singh P, Lee SY, Jeong CH, et al. Isoliquiritigenin induces apoptosis and inhibits xenograft tumor growth of human lung cancer cells by targeting both wild type and L858R/T790M mutant EGFR. *J Bio Chem* 2014; **289**: 35839-48.
- [69] Mishra S, Aeri V, Gaur PK, Jachak SM. Phytochemical, therapeutic, and ethnopharmacological overview for a traditionally important herb: *Boerhavia diffusa* Linn. *BioMed Res Int* 2014; **2014**: 808302. <https://doi.org/10.1155/2014/808302>.
- [70] Cheah KY, Howarth GS, Bindon KA, Kennedy JA, Bastian SE. Low molecular weight procyanidins from grape seeds enhance the impact of 5-Fluorouracil chemotherapy on Caco-2 human colon cancer cells. *PloS One* 2014; **9**: e98921.
- [71] Ali I, Braun DP. Resveratrol enhances mitomycin C-mediated suppression of human colorectal cancer cell proliferation by up-regulation of p21WAF1/CIP1. *Anticancer Res* 2014; **34**: 5439-46.

- [72] Aziz A, Yusran M, Omar AR, Subramani T, Yeap SK, Ho WY, et al. Damcananthal is a potent inducer of apoptosis with anti-cancer activity by stimulating p53 and p21 genes in MCF-7 breast cancer cells. *Oncol Lett* 2014; **7**: 1479-84.
- [73] Guruvayoorappan C, Kuttan G. Immunomodulatory and anti-tumour activity of *Biophytum sensitivium* extract. *Asian Pac J Cancer Prev* 2007; **8**: 27-32.
- [74] Zhan Y, Jia G, Wu D, Xu Y, Xu L. Design and synthesis of a gossypol derivative with improved antitumor activities. *Arch Pharm Chem Life Sci* 2009; **342**: 223-9.
- [75] Shalabi M, Khilo K, Zakaria MM, Elsebaei MG, Abdo W, Awadin W. Anticancer activity of *Aloe vera* and *Calligonum comosum* extracts separately on hepatocellular carcinoma cells. *Asian Pac J Trop Biomed* 2015; **5**(5): 375-81.
- [76] El-Shemy HA, Aboul-Soud MA, Nassr-Allah AA, Aboul-Enein KM, Kabash A, Yagi A, et al. Antitumor properties and modulation of antioxidant enzymes' activity by *Aloe vera* leaf active principles isolated via supercritical carbon dioxide extraction. *Curr Med Chem* 2010; **17**: 129-38.
- [77] Formagio ASN, Vieira MC, Volobuff CRF, Silva MS, Matos AI, Cardoso CAL, et al. *In vitro* biological screening of the anti-cholinesterase and antiproliferative activities of medicinal plants belonging to Annonaceae. *Braz J Med Bio Res* 2015; **48**: 308-15.
- [78] Leyva-Peralta MA, Robles-Zepeda RE, Garibay-Escobar A, Ruiz-Bustos E, Alvarez-Berber LP, Galvez-Ruiz JC, et al. *In vitro* anti-proliferative activity of Argemone gracilenta and identification of some active components. *BMC Comp Altern Med* 2015; **15**: 13.
- [79] Wang Y, Hong C, Zhou C, Xu D, Qu HB. Screening antitumor compounds psoralen and isopsoralen from *Psoralea corylifolia* L. seeds. *Evid-Based Comp Altern Med* 2011; **2011**: 363052. <https://doi.org/10.1093/ecam/nen087>.
- [80] Khalafalla MM, Dafalla HM, Nassrallah A, Aboul-Enein KM, El-Shemy HA, Abdellatef E, et al. Dedifferentiation of leaf explants and antileukemia activity of an ethanolic extract of cell cultures of *Moringa oleifera*. *Afric J Biotech* 2011; **10**: 2746-50.
- [81] Chan LL, George S, Ahmad I, Gosangari SL, Abbasi A, Cunningham BT, et al. Cytotoxicity effects of *Amoora rohituka* and chittagong on breast and pancreatic cancer cells. *Evid base Compl Altern Med* 2011; **2011**: 860605. <https://doi.org/10.1155/2011/860605>.
- [82] Csupor-Löffler B, Hajdu Z, Zupkó I, Molnár J, Forgo P, Vasas A, et al. Antiproliferative constituents of the roots of *Conyza canadensis*. *Plant Med* 2011; **77**: 1183-8.
- [83] Unnati S, Ripal S, Sanjeev A, Niyat A. Novel anticancer agents from plant sources. *Chin J Nat Med* 2013; **11**: 16-23.
- [84] Du GJ, Wang CZ, Qi LW, Zhang ZY, Calway T, He TC, et al. The synergistic apoptotic interaction of panaxadiol and epigallocatechin gallate in human colorectal cancer cells. *Phytother Res* 2013; **27**: 272-7.
- [85] Keglevich P, Hazai L, Kalaus G, Szantay C. Modifications of basic skeleton of vinblastin and vincristine. *Molecules* 2012; **17**: 5893-914.
- [86] Heidari M, Heidari-Vala H, Sadeghi MR, Akhondi MM. The inductive effects of *Centella asiatica* on rat spermatogenic cell apoptosis *in vivo*. *J Nat Med* 2012; **66**: 271-8.
- [87] Bhouri W, Boubaker J, Skandran I, Ghedira K, Ghedira LC. Investigation of the apoptotic way induced by digallic acid in human lymphoblastoid TK6 cells. *Cancer Cell Int* 2012; **12**: 26.
- [88] Raihan MO, Tareq SM, Brishti A, Alam MK, Haque A, Al MS, et al. Evaluation of antitumor activity of *Leea indica* (Burm.f.) merr extract against Ehrlich ascites carcinoma (EAC) bearing mice. *Am J Biomed Sci* 2012; **4**: 143-52.
- [89] Wang L, Xu GF, Liu XX, Chang AX, Xu ML, Ghimeray AK, et al. *In vitro* antioxidant properties and induced G₂/M arrest in HT-29 cells of dichloromethane fraction from *Liriodendron tulipifera*. *J Med Plants Res* 2012; **6**: 424-32.
- [90] Magee PJ, Owusu-Apenten R, McCann MJ, Gill CI, Rowland IR. Chickpea (*Cicer arietinum*) and other plant-derived protease inhibitor concentrates inhibit breast and prostate cancer cell proliferation *in vitro*. *Nutr Cancer* 2012; **64**: 741-8.
- [91] Hoshyar R, Mollaei H. A comprehensive review on anticancer mechanisms of the main carotenoid of saffron, crocin. *J Pharm Pharmacol* 2017. <https://doi.org/10.1111/jphp.12776>.
- [92] Arpita R, Navneeta B. *Centella asiatica*: a pharmaceutically important medicinal plant. *Curr Trends Biomed Eng Biosci* 2017; **5**(3): 555661.
- [93] Kaur R, Karan K, Kaur K. Plants as a source of anticancer agents. *J Nat Prod Plant Resour* 2011; **1**: 119-112.
- [94] Hu M, Xu L, Yin L, Qi Y, Li H, Xu Y, et al. Cytotoxicity of dioscin in human gastric carcinoma cells through death receptor and mitochondrial pathways. *J Appl Toxicol* 2013; **33**: 712-22.
- [95] Motta FCM, Santos DYAC, Salatino MLF, Almeida JMD, Negri G, Carvalho JE, et al. Constituents and antiproliferative activity of extracts from leaves of *Croton macroboothyrs*. *Rev Bras Farmacogn* 2011; **21**: 972-7.
- [96] Maneerat W, Thain S, Cheenpracha S, Prawat U, Laphookhieo S. New amides from the seeds of *Clausana lansium*. *J Med Plants Res* 2011; **5**: 2812-5.
- [97] Lauritano H, Andersen JH, Hansen E, Albrightsen M, Escalera L, Esposito F, et al. Bioactivity screening of microalgae for antioxidant, anti-inflammatory, anticancer, anti-diabetes, and antibacterial activities. *Front Mar Sci* 2016; **3**: 68.
- [98] Hahn ER, Moura MB, Kelley EE, Van Houten B, Shiva S, Singh SV. Withaferin A-induced apoptosis in human breast cancer cells is mediated by reactive oxygen species. *PLoS One* 2011; **6**: e23354.
- [99] Checker R, Sharma D, Sandur SK, Subrahmanyam G, Krishnan S, Poduval TB, et al. Plumbagin inhibits proliferative and inflammatory responses of T cells independent of ROS generation but by modulating intracellular thiols. *J Cell Biochem* 2010; **110**: 1082-93.
- [100] Bakshi HA, Sam S, Anna F, Zeinab R, Ahmad SG, Sharma M, et al. Crocin from Kashmiri saffron (*Crocus sativus*) induces *in vitro* and *in vivo* xenograft growth inhibition of Dalton's lymphoma (DLA) in mice. *Asian Pac J Cancer Prev* 2009; **10**: 887-90.
- [101] Einbond LS, Soffritti M, Esposti DD, Park T, Cruz E, Su T, et al. Actein activates stress-and statin-associated responses and is bioavailable in Sprague-Dawley rats. *Fund Clin Pharmacol* 2009; **23**: 311-21.
- [102] Ogunwande IA, Walker TM, Bansal A, Setzer WN, Essien EE. Essential oil constituents and biological activities of *Peristrophe bicalyculata* and *Borreria verticillata*. *Nat Prod Commun* 2010; **5**: 1815-28.
- [103] Appendino G, Chianese G, Tagliafela-Scalfati O. Cannabinoids: occurrence and medicinal chemistry. *Curr Med Chem* 2011; **18**: 1085-99.
- [104] Colombo V, Lupi M, Falsetta F, Forestieri D, D'Incalci M, Ubezio P, et al. Chemotherapeutic activity of silymarin combined with doxorubicin or paclitaxel in sensitive and multidrug-resistant colon cancer cells. *Canc Chemother Pharmacol* 2011; **67**: 369-79.
- [105] Nakahata AM, Mayer B, Rie C, de Paula CA, Karow M, Neth P, et al. The effects of a plant proteinase inhibitor from *Enterolobium contortisiliquum* on human tumor cell lines. *Biol Chem* 2011; **392**: 327-36.
- [106] Sakarkar DM, Deshmukh VN. Ethnopharmacological review of traditional medicinal plants for anti-cancer activity. *Int J Pharm Technol Res* 2011; **3**: 298-308.
- [107] Xu LN, Lu BN, Hu MM, Xu YW, Han X, Qi Y, et al. Mechanisms involved in the cytotoxic effects of berberine on human colon cancer HCT-8 cells. *Biocell* 2012; **36**: 113-20.
- [108] Karmakar S, Roy Choudhury S, Banik N, Ray S. Molecular mechanisms of anti-cancer action of garlic compounds in neuroblastoma. *Anti Canc Agents Med Chem* 2011; **11**: 398-407.
- [109] Lam SK, Ng TB. Novel galactonic acid-binding hexameric lectin from *Hibiscus mutabilis* seeds with antiproliferative and potent HIV-1 reverse transcriptase inhibitory activities. *Acta Biochim Pol* 2009; **56**: 649.
- [110] Ververidis F, Trantas E, Douglas C, Vollmer G, Kretzschmar G, Panopoulos N, et al. Biotechnology of flavonoids and other phenylpropanoid-derived natural products. Part I: chemical diversity, impacts on plant biology and human health. *Biotechnol J* 2007; **2**: 1214-34.

- [111] Lim do Y, Park JH. Induction of p53 contributes to apoptosis of HCT-116 human colon cancer cells induced by the dietary compound fisetin. *Am J Physiol Gastrointest Liver Physiol* 2009; **296**: 1060-8.
- [112] Madhuri S, Pandey G. Some anticancer medicinal plants of foreign origin. *Curr Sci* 2009; **96**: 6-25.
- [113] Cragg GM, Newman DJ. Plants as a source of anti-cancer agents. *J Ethnopharmacol* 2005; **100**: 72-9.
- [114] Bao MF, Yan JM, Cheng GG, Li XY, Liu YP, Li Y, et al. Cytotoxic indole alkaloids from *Tabernaemontana divaricata*. *J Nat Prod* 2013; **76**: 1406-12.
- [115] Sabnis M. *Chemistry and pharmacology of ayurvedic medicinal plants*. Prakashan: Chaukhamba Amarabharati; 2006.
- [116] Rahman S, Carter P, Bhattacharjee N. Aloe vera for tissue engineering applications. *J Funct Biomater* 2017; **8**(1): 6.
- [117] Agarwal BB, Kumar A, Bharti AC. Anticancer potential of curcumin: preclinical and clinical studies. *Anticancer Res* 2003; **23**: 363-98.
- [118] Merina N, Chandra KJ, Jibon K. Medicinal plants with potential anticancer activities: a review. *Int Res J Phar* 2012; **3**: 26-30.
- [119] Weng JR, Li YB, Chiu CF, Hu JL, Chiu SJ, Wu CY, et al. Cucurbitane triterpenoid from *Momordica charantia* induces apoptosis and autophagy in breast cancer cells, in part, through peroxisome proliferator-activated receptor c activation. *Evid Based Comp Altern Med* 2013; **2013**: 935675. <https://doi.org/10.1155/2013/935675>.
- [120] Kaushik RP, Narayanan V, Vasudevan G, Muthukumaran G, Antony U. Nutrient composition of cultivated *Stevia* leaves and the influence of polyphenols and plant pigments on sensory and antioxidant properties of leaf extracts. *J Food Sci Tech* 2010; **47**: 27-33.
- [121] Das I, Das S, Saha T. Saffron suppresses oxidative stress in DMBA-induced skin carcinoma: a histopathological study. *Acta Histochem* 2010; **112**: 317-27.
- [122] Yoon JS, Kim HM, Yadunandam AK, Kim NH, Jung HA, Choi JS, et al. Neferine isolated from *Nelumbo nucifera* enhances anti-cancer activities in Hep3B cells: molecular mechanisms of cell cycle arrest, ER stress induced apoptosis and anti-angiogenic response. *Phytomed* 2013; **15**: 1013-22.
- [123] Ng TB, Lam YW, Wang H. Calcaelin, a new protein with translation-inhibiting, antiproliferative and antimitogenic activities from the mosaic puffball mushroom *Calvatia caelata*. *Planta Med* 2003; **69**: 212-7.
- [124] Ngai PHK, Ng TB. A ribonuclease with antimicrobial, anti-mitogenic and antiproliferative activities from the edible mushroom *Pleurotus sajor-caju*. *Peptides* 2004; **25**: 11-7.
- [125] Smith JE, Zong A, Rowan NJ. *Medicinal mushrooms: their therapeutic properties and current medical usage with special emphasis on cancer treatments*. London: Canc Res; 2002.
- [126] Srivastava JK, Gupta S. Antiproliferative and apoptotic effects of chamomile extract in various human cancer cells. *J Agric Food Chem* 2007; **55**: 9470-8.
- [127] Alim TK. In: *Edible medicinal and non-medicinal plants*, vol. 5. Netherlands: Springer; 2013. Fruits.
- [128] Suzuki K, Yano T, Sadzuka Y, Sugiyama T, Seki T, Asano R. Restoration of connexin 43 by Bowman-Birk protease inhibitor in M5067 bearing mice. *Oncol Rep* 2005; **13**: 1247-50.
- [129] Huang GJ, Sheu MJ, Chen HJ, Chang YS, Lin YH. Growth inhibition and induction of apoptosis in NB4 promyelocytic leukemia cells by trypsin inhibitor from sweet potato storage roots. *J Agr Food Chem* 2007; **55**: 2548-53.
- [130] Rakashanda S, Mubashir S, Qureshi Y, Hamid A, Masood A, Amin S, et al. Trypsin inhibitors from *Lavatera cashmeriana* camb. seeds: isolation, characterization and *in-vitro* cytotoxic activity. *Int J Pharm Sci Invent* 2013b; **2**: 55-65.
- [131] Caccialupi P, Ceci LR, Siciliano RA, Pignone D, Clemente A, Sonnante G. Bowman-birk inhibitors in lentil: heterologous expression, functional characterisation and anti-proliferative properties in human colon cancer cells. *Food Chem* 2010; **120**: 1058-66.
- [132] Lanza A, Tava A, Catalano M, Ragona L, Singuarioli I, Robustelli della Cuna FS, et al. Effects of the *Medicago scutellata* trypsin inhibitor (MsTI) on cisplatin-induced cytotoxicity in human breast and cervical cancer cells. *Anticancer Res* 2004; **24**: 227-34.
- [133] Sun J, Wang H, Ng TB. Trypsin isoforms with antiproliferative activity toward leukemia cells from *Phaseolus vulgaris* cv 'white cloud bean'. *J Biomed Biotech* 2010; **2010**: 219793.
- [134] Runchana R, Wanee J, Pea, *Pisum sativum*, and its anticancer activity. *Pharmacog Rev* 2017; **11**(21): 39-42.
- [135] Sathesh LS, Murugan K. Antimicrobial activity of protease inhibitor from leaves of *Coccinia grandis* (L.) Voigt. *Ind J Exp Biol* 2011; **49**: 366-74.
- [136] Suzuki R, Kohno H, Sugie S, Sasaki K, Yoshimura T, Wada K, et al. Preventive effects of extract of leaves of ginkgo (*Ginkgo biloba*) and its component bilobalide on azoxymethane-induced colonic aberrant crypt foci in rats. *Canc Lett* 2004; **210**: 159-69.
- [137] Seo WG, Hwang JC, Kang SK, Jin UH, Suh SJ, Moon SK, et al. Suppressive effect of *Zedoariae rhizoma* on pulmonary metastasis of B16 melanoma cells. *J Ethnopharmacol* 2005; **101**: 249-57.
- [138] Zhao Y, Wang CM, Wang BG, Zhang CX. Study on the anti-cancer activities of the *Clematis manshurica* saponins in vivo. *China J Chin Mater Med* 2005; **30**: 1452-3.
- [139] Imai M, Kikuchi H, Denda T, Ohyama K, Hirobe C, Toyoda H, et al. Cytotoxic effects of flavonoids against a human colon cancer derived cell line, COLO 201: a potential natural anti-cancer substance. *Canc Lett* 2009; **276**: 74-80.
- [140] Yadav B, Bajaj A, Saxena M, Saxena AK. *In vitro* anticancer activity of the root, stem and leaves of *Withania somnifera* against various human cancer cell lines. *Indian J Pharm Sci* 2010; **72**: 659.
- [141] Benarba B, Meddah B. Ethnobotanical study, antifungal activity, phytochemical screening and total phenolic content of Algerian *Aristolochia longa*. *J Intercult Ethnopharmacol* 2014; **3**: 150.
- [142] Babu S, Padikkala J, Sathiadevan P, Vijayakurup V, Azis T, Srinivas P, et al. Apoptosis induction of *Centella asiatica* on human breast cancer cells. *Afr J Tradit Complement Altern Med* 2009; **25**: 9-16.
- [143] Sahreen S, Khan MR, Khan RA, Shah NA. Estimation of flavonoids, antimicrobial, antitumor and anticancer activity of *Carissa opaca* fruits. *BMC Compl Alter Med* 2013; **13**: 372.
- [144] Li J, Li QC, Chen C, Hao X, Liu H. Cytotoxic activity of cardenolides and cardenolide glycosides from *Asclepias curassavica*. *Bioorganic Med Chem Lett* 2009; **19**: 1956-9.
- [145] Biba VS, Jeba MPW, Remani P. Differential effects of *Annona squamosa* seed extracts: antioxidant, antibacterial, cytotoxic and apoptotic study. *Int J Pharm Biol Sci* 2013; **4**: 899-907.
- [146] Mahata S, Maru S, Shukla S, Pandey A, Mugesh G, Das BC, et al. Anticancer property of *Bryophyllum pinnata* (Lam.) Oken. leaf on human cervical cancer cells. *BMC Compl Alter Med* 2012; **12**: 15.
- [147] Choedon T, Shukla SK, Kumar V. Chemopreventive and anti-cancer properties of the aqueous extract of flowers of *Butea monosperma*. *J Ethnopharmacol* 2010; **129**: 208-13.
- [148] Awale S, Linn TZ, Li F. Identification of chrysoplenetin from *Vitex negundo* as a potential cytotoxic agent against PANC-1 and a panel of 39 human cancer cell lines (JFCR39). *Phytother Res* 2011; **25**: 1770-5.
- [149] Shaban A, Mani GM, Nautiyal R. *In vitro* cytotoxicity of *Moringa oleifera* against different human cancer cell lines. *Asian J Pharma Clin Res* 2012; **5**: 271-2.
- [150] Afify MRA, Fayed AS, Shalaby EA, El-Shemy AH. *Syzygium cumini* (pomposia) active principles exhibit potent anticancer and antioxidant activities. *Afr J Pharm Pharmacol* 2011; **5**: 948-56.
- [151] Brahmachari G, Gorai D, Roy R. Argemone mexicana: chemical and pharmacological aspects. *Braz J Pharmacog* 2013; **23**: 559-75.
- [152] Hirata T, Fujii M, Akita K. Identification and physiological evaluation of the components from Citrus fruits as potential drugs for anti-corpulence and anticancer. *Bioorgan Med Chem* 2009; **17**: 25-8.
- [153] Reddy KP, Bid HK, Nayak VL. *In-vitro* and *in-vivo* anticancer activity of 2-deacetoxytaxinine J and synthesis of novel taxoids and their *in vitro* anticancer activity. *Eur J Med Chem* 2009; **44**: 3947-53.
- [154] aDe Bono JS, Oudard S, Ozguroglu M, Hansen S, Machiels JP, Kocak I, et al. Prednisone plus cabazitaxel or mitoxantrone for metastatic castration-resistant prostate cancer progressing after docetaxel treatment: a randomised open-label trial. *Lancet* 2010; **376**: 1147-54.

- [155] Atkinson JM, Falconer RA, Edwards DR, Pennington CJ, Siller CS, Shnyder SD, et al. Development of a novel tumor-targeted vascular disrupting agent activated by membrane-type matrix metalloproteinases. *Canc Res* 2010; **70**: 6902-12.
- [156] Dieras V, Limentani S, Romieu G, Tubiana-Hulin M, Lortholary A, Kaufman P, et al. Phase II multicenter study of larotaxel (XRP9881), a novel taxoid, in patients with metastatic breast cancer who previously received taxane-based therapy. *Ann Oncol* 2008; **19**(7): 1255-60.
- [157] Pierpaoli E, Damiani E, Orlando F, Lucarini G, Bartozzi B, Lombardi P, et al. Antiangiogenic and antitumor activities of berberine derivative NAX014 compound in a transgenic murine model of HER2/neu-positive mammary carcinoma. *Carcinogenesis* 2015; **36**(10): 1169-79.
- [158] Singh S, Jarial R, Kanwar SS. Therapeutic effect of herbal medicines on obesity: herbal pancreatic lipase inhibitors. *Wud-pecker J Med Plants* 2013; **2**: 53-65.
- [159] Maryam M, Go R, Yien CYS, Nazre M. Vinca alkaloids. *Int J Prev Med* 2013; **4**: 1231-5.
- [160] Almagro L, Fernández-Perez F, Pedreno MA. Indole alkaloids from *Catharanthus roseus*: bioproduction and their effect on human health. *Molecules* 2015; **20**: 2973-3000.
- [161] Xie S, Zhou J. Harnessing plant biodiversity for the discovery of novel anticancer drugs targeting microtubules. *Front Plant Sci* 2017; **8**: 720.
- [162] Schutz FA, Bellmunt J, Rosenberg JE, Choueiri TK. Vinflunine: drug safety evaluation of this novel synthetic vinca alkaloid. *Expert Opin Drug Saf* 2011; **10**: 645-53.
- [163] Darshan MS, Loftu MS, Thadani-Mulero M, Levy BP, Escuin D, Zhou XK, et al. Taxane-induced blockade to nuclear accumulation of the androgen receptor predicts clinical responses in metastatic prostate cancer. *Cancer Res* 2011; **71**: 6019-29.
- [164] Ojima I, Lichtenthal B, Lee S, Wang C, Wang X. Taxane anticancer agents: a patent perspective. *Expert Opinon Ther Pat* 2016; **26**: 1-20.
- [165] Kim SH, Kaplan JA, Sun Y, Shieh A, Sun HL, Croce CM, et al. The self-assembly of anticancer camptothecin-dipeptide nanotubes: a minimalistic and high drug loading approach to increased efficacy. *Chem-A Eurp J* 2015; **21**: 101-5.
- [166] Rahier NJ, Thomas CJ, Hecht SM. Camptothecin and its analogs. In: Cragg GM, Kingston DGI, New DG, editors. *Anticancer agents from natural products*. Boca Raton, FL: Taylor and Francis; 2005, p. 5-22.
- [167] Heibligha M, Sobha M, Nicolini FE. Subcutaneous omacetaxine mepesuccinate in patients with chronic myeloid leukemia in tyrosine kinase inhibitor-resistant patients. *Rev Pers* 2014; **38**: 1145-53.
- [168] Feldman EJ, Seiter KP, Ahmed T, Baskind P, Arlin ZA. Homoharringtonine in patients with myelodysplastic syndrome (MDS) and MDS evolving to acute myeloid leukemia. *Leukemia* 1996; **10**: 40-2.
- [169] Li FS, Weng JK. Demystifying traditional herbal medicine with modern approaches. *Nat Plants* 2017; **3**(8): 1-7.
- [170] Negia AS, Gautama Y, Alama S, Chanda D, Luqmana S, Sarkar J, et al. Natural antitubulin agents: Importance of 3,4,5-trimethoxyphenyl fragment. *Bioorg Med Chem* 2015; **23**: 373-89.
- [171] Stiborova M, Cerna V, Moserova M, Mrzova I, Arlt VM, Frei E, et al. The anticancer drug ellipticine activated with cytochrome P450 mediates DNA damage determining its pharmacological efficiencies: studies with rats, hepatic cytochrome P450 reductase null (HRNTM) mice and pure enzymes. *Int J Mol Sci* 2015; **16**: 284-306.
- [172] Kizek R, Adam V, Hrabeta J, Eckschlager T, Smutny S, Burda JV. Anthracyclines and ellipticines as DNA-damaging anticancer drugs: recent advances. *Phar Ther* 2012; **13**: 26-39.
- [173] Isah T. Anticancer alkaloids from trees: development into drugs. *Phcog Rev* 2016; **10**: 90-9.
- [174] Ohashi M, Sugikawa E, Nakanishi N. Inhibition of p53 protein phosphorylation by 9-hydroxyellipticine: a possible anticancer mechanism. *Jpn J Cancer Res* 1995; **86**: 819-29.
- [175] Mantena SK, Sharma SD, Katiyar SK. Berberine, a natural product, induces G1-phase cell cycle arrest and caspase-3-dependent apoptosis in human prostate carcinoma cells. *Mol Cancer Ther* 2006; **5**: 296-308.
- [176] Barzegar E, Fouladdel SH, Movahhed TK, Atashpour SH, Ghahremani HM, Ostad SN, et al. Effects of berberine on proliferation, cell cycle distribution and apoptosis of human breast cancer T47D and MCF7 cell lines. *Iran J Basic Med Sci* 2015; **18**: 334-42.
- [177] Xu H, Zhang S. Scutellarin induced apoptosis in HepG2 hepatocellular carcinoma cells via a STAT3 pathway. *Phytother Res* 2013; **27**(10): 1524-8.
- [178] Garon EB, Neidhart JD, Gabrail NY, de Oliveira MR, Balkissoon J, Kabbinavar F. A randomized phase II trial of the tumor vascular disrupting agent CA4P (fosbretabulin tromethamine) with carboplatin, paclitaxel, and bevacizumab in advanced nonsquamous non-small-cell lung cancer. *Oncotargets Ther* 2016; **9**: 7275.
- [179] Prakash O, Kumar A, Kumar P. Anticancer potential of plants and natural products: a review. *Am J Pharmacol Sci* 2013; **1**: 104-15.
- [180] Venier NA, Yamamoto T, Sugar LM, Adomat H, Fleshner NE, Klotz LH, et al. Capsaicin reduces the metastatic burden in the transgenic adenocarcinoma of the mouse prostate model. *Prostate* 2015; **75**(12): 1300-11.
- [181] Clark R, Lee SH. Anticancer properties of capsaicin against human cancer. *Anticancer Res* 2016; **36**(3): 837-43.
- [182] Chang HC, Chen ST, Chien SY, Kuo SJ, Tsai HT, Chen DR. Capsaicin may induce breast cancer cell death through apoptosis-inducing factor involving mitochondrial dysfunction. *Hum Exp Toxicol* 2011; **30**(10): 1657-65.
- [183] Sarkar A, Bhattacharjee S, Mandal DP. Induction of apoptosis by eugenol and capsaicin in human gastric cancer AGS cells: elucidating the role of p53. *Asian Pac J Cancer Prev* 2015; **16**(16): 6753-9.
- [184] Kim S, Moon A. Capsaicin-Induced apoptosis of h-ras-transformed human breast epithelial cells is rac-dependent via ras generation. *Arch Pharm Res* 2004; **27**(8): 845-9.
- [185] Mallory SR, Zwick JC, Pei P, Tong M, Larsen PE, Shumway BS, et al. Topical application of a bioadhesive black raspberry gel modulates gene expression and reduces cyclooxygenase 2 protein in human premalignant oral lesions. *Cancer Res* 2008; **68**: 4945-57.
- [186] Wang LS, Burke CA, Hasson H, Kuo CT, Molmenti CLS, Seguin C, et al. A phase Ib study of the effects of black raspberries on rectal polyps in patients with familial adenomatous polyposis. *Cancer Prev Res* 2014; **7**: 666-74.
- [187] Chen J, Mu Q, Li X, Yin X, Yu M, Jin J, et al. Homoharringtonine targets Smad3 and TGF- β pathway to inhibit the proliferation of acute myeloid leukemia cells. *Oncotarget* 2017; **8**(25): 40318.
- [188] Rathkopf D, Dickson MA, Feldman DR, Carvajal RD, Shah MA, Wu N, et al. Phase I study of flavopiridol with oxaliplatin and fluorouracil/leucovorin in advanced solid tumors. *Clin Cancer Res* 2009; **15**: 7405-11.
- [189] Morimoto Y, Maskarinec G, Park SY, Ettienne R, Matsuno RK, Long C, et al. Dietary isoflavone intake is not statistically significantly associated with breast cancer risk in the multiethnic cohort. *Br J Nutr* 2014; **112**: 976-83.
- [190] Tang NP, Zhou B, Wang B, Yu RB, Ma J. Flavonoids intake and risk of lung cancer: a meta-analysis. *Jpn J Clin Oncol* 2009; **39**: 352-9.
- [191] Woo HD, Kim J. Dietary flavonoid intake and risk of stomach and colorectal cancer. *World J Gastroenterol* 2013; **19**: 1011-9.
- [192] Kim JE, Kwon JY, Seo SK, Son JE, Jung SK, Min SY, et al. Cyanidin suppresses ultraviolet B-induced COX-2 expression in epidermal cells by targeting MKK4, MEK1, and Raf-1. *Biochem Pharmacol* 2010; **79**: 1473-82.
- [193] Singh BN, Shankar S, Srivastava RK. Green tea catechin, epigallocatechin-3-gallate (EGCG): mechanisms, perspectives and clinical applications. *Biochem Pharmacol* 2011; **82**: 1807-21.
- [194] Luthra PM, Lal N. Prospective of curcumin, a pleiotropic signaling molecule from Curcuma longa in the treatment of glioblastoma. *Eur J Med Chem* 2016; **109**: 23-35.
- [195] Patel S, Sarwat M, Khan TH. Mechanism behind the anti-tumour potential of saffron (*Crocus sativus* L.): the molecular perspective. *Crit Rev Oncol/Hematolo* 2017; **115**: 27-35.

- [196] Morris J, Moseley VR, Cabang AB, Coleman K, Wei W, Garrett-Mayer E, et al. Reduction in promotor methylation utilizing EGCG (Epigallocatechin-3-gallate) restores RXR α expression in human colon cancer cells. *Oncotarget* 2016; **7**: 11-7.
- [197] Qiao Y, Cao J, Xie L, Shi X. Cell growth inhibition and gene expression regulation by (-)-epigallocatechin-3-gallate in human cervical cancer cells. *Arch Pharm Res* 2009; **32**: 1309-15.
- [198] Budisan L, Gulei D, Zanoaga OM, Irimie AI, Sergiu C, Braicu C, et al. Dietary Intervention by phytochemicals and their role in modulating coding and non-coding genes in cancer. *Int J Mol Sci* 2017; **18**(6): 1178.
- [199] Philips BJ, Coyle CH, Morrisroe SN, Chancellor MB, Yoshimura N. Induction of apoptosis in human bladder cancer cells by green tea catechins. *Biomed Res* 2009; **30**: 207-15.
- [200] Park YJ, Wen J, Bang S, Park SW, Song SY. [6]-Gingerol induces cell cycle arrest and cell death of mutant p53-expressing pancreatic cancer cells. *Yonsei Med J* 2006; **47**: 688-97.
- [201] Oyagbemi A, Saba A, Azeem O. Capsaicin: a novel chemopreventive molecule and its underlying molecular mechanisms of action. *Indian J Cancer* 2010; **47**: 53-8.
- [202] Bernard MM, McConnery JR, Hoskin DW. [10]-Gingerol, a major phenolic constituent of ginger root, induces cell cycle arrest and apoptosis in triple-negative breast cancer cells. *Exp Mol Pathol* 2017; **102**(2): 370-6.
- [203] Nahum A, Hirsch K, Danilenko M, Watts C, Prall O, Levy J, et al. Lycopene inhibition of cell cycle progression in breast and endometrial cancer cells is associated with reduction in cyclin D levels and retention of p27 (Kip1) in the cyclin E-cdk2 complexes. *Oncogene* 2001; **20**: 3428-36.
- [204] Carini F, David S, Tomasello G, Mazzola M, Damiani P, Rappa F, et al. Colorectal cancer: an update on the effects of lycopene on tumor progression and cell proliferation. *JBRHA* 2017; **31**(3): 1-9.
- [205] Cha JH, Kim WK, Ha AW, Kim MH, Chang MJ. Anti-inflammatory effect of lycopene in SW480 human colorectal cancer cells. *Nutr Res Pract* 2017; **11**(2): 90-6.
- [206] Gorham ED, Garland CF, Garland FC, Grant WB, Mohr SB, Lipkin M. Optimal vitamin D status for colorectal cancer prevention: a quantitative meta-analysis. *Am J Prev Med* 2007; **32**: 210-6.
- [207] Buyru N, Tezol A, Yosunkaya-Fenerci E, Dalay N. Vitamin D receptor gene polymorphisms in breast cancer. *Exp Mol Med* 2003; **35**: 550-5.
- [208] Garland CF, Mohr SB, Gorham ED, Grant WB, Garland FC. Role of ultraviolet B irradiance and vitamin D in prevention of ovarian cancer. *Am J Prev Med* 2006; **31**: 512-4.
- [209] Zong A, Cao H, Wang F. Anticancer polysaccharides from natural resources: a review of recent research. *Carbohydr Polym* 2012; **90**: 1395-410.
- [210] Maehara Y, Tsujitani S, Saeki H. Biological mechanism and clinical effect of protein-bound polysaccharide K (KRESTIN): review of development and future perspectives. *Surg Today* 2012; **42**: 8-28.
- [211] Ivanova TS, Krupodorova TA, Barshteyn VY, Artamonova AB, Shlyakhovenko VA. Anticancer substances of mushroom origin. *Exp Oncol* 2014; **36**: 58-66.
- [212] Mikashvili N, Elisashvili V, Worku M. Purification and characterization of a lectin isolated from the submerged cultivated mycelium of grey polypore *Cerrena unicolor* (Bull.) Murrill (Aphyllonomycetidae). *Int J Med Mushr* 2009; **11**: 61-8.
- [213] Xu W, Huang JJ, Cheung PC. Extract of *Pleurotus pulmonarius* suppresses liver cancer development and progression through inhibition of VEGF-induced PI3K/AKT signaling pathway. *PLoS One* 2011; **7**: e34406.
- [214] Barbieri A, Quagliariello V, Del Vecchio V, Falco M, Luciano A, Amruthraj, et al. Anticancer and anti-inflammatory properties of ganoderma lucidum extract effects on melanoma and triple-negative breast cancer treatment. *Nutrients* 2017; **9**(3): 210.
- [215] Awadasseid A, Hou J, Gamallat Y, Xueqi S, Eugene KD, Hago AM, et al. Purification, characterization, and antitumor activity of a novel glucan from the fruiting bodies of *Coriolus versicolor*. *PLoS One* 2017; **12**(2): e0171270.
- [216] Wada S. Cancer preventive effects of vitamin E. *Curr Pharm Biotechnol* 2012; **13**: 156-64.
- [217] Geraets L, Haegens A, Brauers K, Haydock JA, Vernooy JH, Wouters EF, et al. Inhibition of LPS-induced pulmonary inflammation by specific flavonoids. *Biochem Biophys Res Comm* 2009; **382**: 598-603.
- [218] Khan N, Afaq F, Khusro FH, Adhami VM, Suh Y, Mukhtar H. Dual inhibition of PI3K/AKT and mTOR signaling in human non-small cell lung cancer cells by a dietary flavonoid fisetin. *Int J Canc* 2012; **130**: 1695.
- [219] Min KJ, Nam JO, Kwon TK. Fisetin induces apoptosis through p53-mediated up-regulation of DR5 expression in human renal carcinoma caki cells. *Molecule* 2017; **22**(8): 1285.
- [220] Patel KR, Brown VA, Jones DJ, Britton RG, Hemingway D, Miller AS, et al. Clinical pharmacology of resveratrol and its metabolites in colorectal cancer patients. *Canc Res* 2010; **70**: 7392-9.
- [221] Varoni EM, Faro AFL, Sharifi-Rad J, Iriti M. Anticancer molecular mechanisms of resveratrol. *Front Nutr* 2016; **3**: 8.
- [222] Brakenhielm E, Cao R, Cao Y. Suppression of angiogenesis, tumor growth, and wound healing by resveratrol, a natural compound in red wine and grapes. *FASEB J* 2001; **15**(10): 1798-800.
- [223] Jaswanth A, Vasanthi HR, Rajamanickam GV, Saraswathy A. Tumoricidal effect of the red algae *Acanthophora spicifera* on Ehrlich's ascites carcinoma cellsutilization. *Seaweed Res* 2004; **26**(1-2): 217-23.
- [224] Yuan YV, Carrington MF, Walsh NA. Extracts from dulse (*Palmaria palmata*) are effective antioxidants and inhibitors of cell proliferation in vitro. *Food Chem Toxicol* 2005; **43**: 1073-81.
- [225] Nuhu AA. Spirulina (*Arthrospira*): an important source of nutritional and medicinal compounds. *J Mar Biol* 2013; **2013**: 325636. <https://doi.org/10.1155/2013/325636>.
- [226] Seo HS, Jo JK, Ku JM, Choi HS, Choi YK. Induction of caspase dependent extrinsic apoptosis through inhibition of signal transducer and activator of transcription 3 (STAT3) signaling in HER2-overexpressing BT-474 breast cancer cells. *Biosci Rep* 2015; **35**: e00276.
- [227] Bauer D, Redmon N, Mazzio E, Soliman KF. Apigenin inhibits TNF α /IL-1 α -induced CCL2 release through IKBK-epsilon signaling in MDA-MB-231 human breast cancer cells. *PLoS One* 2017; **12**(4): e0175558.
- [228] Liu J, Zhang Y, Qu J, Xu L, Hou K, Zhang J, et al. β -Elemene-induced autophagy protects human gastric cancer cells from undergoing apoptosis. *BMC Cancer* 2011; **11**: 183.
- [229] Jiang Z, Jacob JA, Loganathachetti DS, Nainangu P, Chen B. β -elemene: mechanistic studies on cancer cell interaction and its chemosensitization effect. *Front Pharmacol* 2017; **8**: 105.
- [230] Jiang S, Ling C, Li W, Jiang H, Zhi Q, Jiang M. Molecular mechanisms of anti-cancer activities of β -elemene: targeting hallmarks of cancer. Anti-cancer agents in medicinal chemistry. *Form Curr Med Chem AntiCancer Agent* 2016; **16**: 1426-34.
- [231] Das M, Manna K. Chalcone scaffold in anticancer armamentarium: a molecular insight. *J Toxicol* 2016; **2016**: 7651047. <https://doi.org/10.1155/2016/7651047>.
- [232] Shaobi M, Shah I, Ali N, Adhikari A, Tahir MN, Shah SWA. Sesquiterpene lactone! a promising antioxidant, anticancer and moderate antinociceptive agent from *Artemisia macrocephala jacquem*. *BMC Compl Altern Med* 2017; **17**(1): 27.
- [233] Mehmood T, Maryam A, Ghram HA, Khan M, Ma T. Deoxyelephantopin and isodeoxyelephantopin as potential anticancer agents with effects on multiple signaling pathways. *Molecules* 2017; **22**(6): 1013.
- [234] Kasala ER, Bodduluru LN, Madana R, Gogoi R, Barua CC. Chemopreventive and therapeutic potential of chrysin in cancer: mechanistic perspectives. *Toxicol Lett* 2015; **233**: 214-25.
- [235] Renuka M, Vijayakumar N, Ramakrishnan A. Chrysin, a flavonoid attenuates histological changes of hyperammonemic rats: a dose dependent study. *Biomed Pharmacother* 2016; **82**: 345-54.
- [236] Ryu S, Lim W, Bazer FW, Song G. Chrysin induces death of prostate cancer cells by inducing ROS and ER stress. *J Cell Physiol* 2017; **232**(12): 3786-97.

- [237] Xing J, You H, Dong Y, Lu J, Chen S, Zhu H, et al. Metabolic and pharmacokinetic studies of scutellarin in rat plasma, urine, and feces. *Acta Pharmacol Sin* 2011; **32**: 655-63.
- [238] Gao C, Zhou Y, Jiang Z, Zhao Y, Zhang D, Cong X, et al. Cytotoxic and chemosensitization effects of Scutellarin from traditional Chinese herb *Scutellaria altissima* L. in human prostate cancer cells. *Oncol Rep* 2017; **38**(3): 1491-9.
- [239] Yang N, Zhao Y, Wang Z, Liu Y, Zhang Y. Scutellarin suppresses growth and causes apoptosis of human colorectal cancer cells by regulating the p53 pathway. *Mol Med Rep* 2017; **15**(2): 929-35.
- [240] Chen YC, Yang LL, Lee TJF. Oroxylin A inhibition of lipopolysaccharide-induced iNOS and COX-2 gene expression via suppression of nuclear factor- κ B activation. *Biochem Pharmacol* 2004; **59**: 1445-57.
- [241] Ha J, Zhao L, Zhao Q, Yao J, Zhu BB, Lu N, et al. Oroxylin A improves the sensitivity of HT-29 human colon cancer cells to 5-FU through modulation of the COX-2 signaling pathway. *Biochem Cell Biol* 2012; **90**: 521-31.
- [242] Lee HS, Cho HJ, Yu R, Lee KW, Chun HS, Park JHY. Mechanisms underlying apoptosis-inducing effects of kaempferol in HT-29 human colon cancer cells. *Int J Mol Sci* 2014; **15**: 2722-37.
- [243] Gutiérrez-del-Río I, Villar CJ, Lombó F. Therapeutic uses of kaempferol: anticancer and anti-inflammatory activity. In: Garde-Cerdán T, Gonzalo-Diago A, editors. *Kaempferol: Biosynthesis, food sources and therapeutic uses*. New York: Nova Science Publishers; 2016.
- [244] Lee GA, Choi KC, Hwang KA. Kaempferol, a phytoestrogen, suppressed tricosan-induced epithelial-mesenchymal transition and metastatic-related behaviors of MCF-7 breast cancer cells. *Environ Toxicol Pharmacol* 2017; **49**: 48-57.
- [245] Marin L, Miguelez EM, Villar CJ, Lombó F. Bioavailability of dietary polyphenols and gut microbiota metabolism: antimicrobial properties. *Biomed Res Int* 2015; **1120**: 905215.
- [246] Mizushina Y, Shiomi K, Kuriyama I, Takahashi Y, Yoshida H. Inhibitory effects of a major soy isoflavone, genistein, on human DNA topoisomerase II activity and cancer cell proliferation. *Int J Oncol* 2013; **43**: 1117-24.
- [247] Luo Y, Wang SX, Zhou ZQ, Wang Z, Zhang YG, Zhang Y, et al. Apoptotic effect of genistein on human colon cancer cells via inhibiting the nuclear factor- κ B (NF- κ B) pathway. *Tumour Biol* 2014; **35**: 11483-8.
- [248] Ganai AA, Farooqi H. Bioactivity of genistein: a review of *in vitro* and *in vivo* studies. *Biomed Pharmacoth* 2015; **76**: 30-8.
- [249] Lee JI, Hsu BH, Wu D, Barrett JS. Separation and characterization of silybin, isosilybin, silydianin and silychristin in milk thistle extract by liquid chromatography electrospray tandem mass spectrometry. *J Chromatogr A* 2006; **1116**: 57-68.
- [250] Thorn CF, Oshiro C, Marsh S, Hernandez-Boussard T, McLeod H, Klein TE, et al. Doxorubicin pathways: pharmacodynamics and adverse effects. *Pharmacogenet Genomics* 2011; **21**: 440-6.
- [251] Kim SH, Ryu HG, Lee J, Shin J, Harikishore A, Jung HY, et al. Ursolic acid exerts anti-cancer activity by suppressing vaccinia-related kinase 1-mediated damage repair in lung cancer cells. *Sci Rep* 2015; **5**: 14570.
- [252] Chen XJ, Zhang XJ, Shui YM, Wan JB, Gao JL. Anticancer activities of protopanaxadiol-and protopanaxatriol-type ginsenosides and their metabolites. *Evid Based Complement Altern Med* 2016; **2016**: 1-19.
- [253] Ahuja A, Kim JH, Kim JH, Yi YS, Cho JY. Functional role of ginseng-derived compounds in cancer. *J Ginseng Res* 2017; **16**: 1-7.
- [254] Gao JL, Lv GY, He BC, Zhang BQ, Zhang H, Wang N, et al. Ginseng saponin metabolite 20(S)-protopanaxadiol inhibits tumor growth by targeting multiple cancer signaling pathways. *Oncol Rep* 2013; **30**: 292-8.
- [255] Kim SM, Lee SY, Yuk DY, Moon DC, Choi SS, Kim Y, et al. Inhibition of 1043 NF- κ B by ginsenoside Rg3 enhances the susceptibility of colon cancer cells to docetaxel. *Arch Pharm Res* 2009; **32**: 755-65.
- [256] Liang LD, He T, Du TW, Fan YG, Chen DS, Wang Y. Ginsenoside-Rg5 induces apoptosis and DNA damage in human cervical cancer cells. *Mol Med Rep* 2015; **11**(2): 940-6.
- [257] Zhu H, Ding WJ, Wu R, Weng QJ, Lou JS, Jin RJ, et al. Synergistic anti-cancer activity by the combination of TRAIL/APO-2L and celastrol. *Cancer Invest* 2010; **28**: 23-32.
- [258] Yu X, Ruan X, Zhang J, Zhao Q. Celastrol induces cell apoptosis and inhibits the expression of the AML1-ETO/C-KIT oncogene in t (8; 21) leukemia. *Molecules* 2016; **21**(5): 574.
- [259] Fan XX, Li N, Wu JL, Zhou YL, He JX, Liu L, et al. Celastrol induces apoptosis in gefitinib-resistant non-small cell lung cancer cells via caspases-dependent pathways and Hsp90 client protein degradation. *Molecules* 2014; **19**(3): 3508-22.
- [260] Cascao R, Fonseca JE, Moita LF. Celastrol: a spectrum of treatment opportunities in chronic diseases. *Front Med* 2017; **4**: 69.
- [261] Meng X, Liang H, Luo L. Antitumor polysaccharides from mushrooms: a review on the structural characteristics, antitumor mechanisms and immunomodulating activities. *Carbohydr Res* 2016; **424**: 30-41.
- [262] Schepetkin IA, Quinn MT. Botanical polysaccharides: macrophage immunomodulation and therapeutic potential. *Int Immunopharmacol* 2006; **6**: 317-33.
- [263] Zhang Y, Li Q, Wang J, Cheng F, Huang X, Cheng Y, et al. Polysaccharide from *Lentinus edodes* combined with oxaliplatin possesses the synergy and attenuation effect in hepatocellular carcinoma. *Cancer Lett* 2016; **377**: 117-25.
- [264] Chu BF, Lin HC, Huang XW, Huang HY, Wu CP, Kao MC, et al. An ethanol extract of *Poria cocos* inhibits the proliferation of non-small cell lung cancer A549 cells via the mitochondria-mediated caspase activation pathway. *J Funct Foods* 2016; **23**: 614-27.
- [265] Liu G, Kuang S, Wu S, Jin W, Sun C. A novel polysaccharide from *Sargassum integrerrimum* induces apoptosis in A549 cells and prevents angiogenesis *in vitro* and *in vivo*. *Sci Rep* 2016; **6**: 26722.
- [266] Sun Q, Dong M, Wang Z, Wang C, Sheng D, Li Z, et al. Selenium enriched polysaccharides from *Pyracantha fortuneana* (Se-PFPs) inhibit the growth and invasive potential of ovarian cancer cells through inhibiting beta catenins signaling. *Oncotarget* 2016; **7**: 28369-83.
- [267] Luo B, Wang J, Li X, Lu W, Yang J, Hu Y, et al. New mild and simple approach to isothiocyanates: a class of potent anticancer agents. *Molecules* 2017; **22**(6): 773.
- [268] Pereira LP, Silva P, Duarte M, Rodrigues L, Duarte CM, Albuquerque C, et al. Targeting colorectal cancer proliferation, stemness and metastatic potential using brassicaceae extracts enriched in isothiocyanates: a 3D cell model-based study. *Nutrients* 2017; **9**(4): 368.
- [269] Cheng YM, Tsai CC, Hsu YC. Sulforaphane, a dietary isothiocyanate, induces G₂/M arrest in cervical cancer cells through cyclinB1 downregulation and GADD45 β /CDC2 association. *Int J Mol Sci* 2016; **17**(9): 1530.
- [270] Shindo S, Hosokawa Y, Hosokawa I, Ozaki K, Matsuo T. Genipin inhibits MMP-1 and MMP-3 release from TNF- α -stimulated human periodontal ligament cells. *Biochimie* 2014; **107**: 391-5.
- [271] Pons DG, Nadal-Serrano M, Torrens-Mas M, Valle A, Oliver J, Roca P. UCP2 inhibition sensitizes breast cancer cells to therapeutic agents by increasing oxidative stress. *Free Radic Biol Med* 2015; **86**: 67-77.
- [272] Liu HE, Chang ASY, Teng CM, Chen CC, Tsai AC, Yang CR. Potent anti-inflammatory effects of denbinobin mediated by dual inhibition of expression of inducible nitric oxide synthase and cyclooxygenase 2. *Shock* 2011; **35**(2): 191-7.
- [273] Peiro G, Ortiz-Martinez F, Gallardo A, Perez-Balaguer A, Sanchez-Paya J, Ponce JJ, et al. Src, a potential target for overcoming trastuzumab resistance in HER2-positive breast carcinoma. *Br J Cancer* 2014; **111**(4): 689.